Real-time monitoring of nutrients and dissolved organic matter in rivers: capturing event
dynamics, technological opportunities and future directions

Authors:
Phillip J. Blaen\textsuperscript{a,b}
Kieran Khamis\textsuperscript{a}
Charlotte E.M. Lloyd\textsuperscript{c}
Chris Bradley\textsuperscript{a}
David Hannah\textsuperscript{a}
Stefan Krause\textsuperscript{a}

\textsuperscript{a} School of Geography, Earth and Environmental Sciences, University of Birmingham, Edgbaston, Birmingham, B15 2TT, UK
\textsuperscript{b} Birmingham Institute of Forest Research (BIFoR), University of Birmingham, Edgbaston, Birmingham, B15 2TT, UK
\textsuperscript{c} Organic Geochemistry Unit, Bristol Biogeochemistry Research Centre, School of Chemistry, University of Bristol, Cantocks Close, Bristol, BS8 1TS, UK

Corresponding author:
Phillip J. Blaen
p.j.blaen@bham.ac.uk

Keywords:
In-situ, high-frequency, nitrate, phosphate, DOC, DOM, sensor
Abstract

Excessive riverine nutrient concentrations threaten aquatic ecosystem structure and functioning and can pose substantial risks to human health. Robust monitoring strategies are therefore required to generate reliable estimates of river nutrient loads and to improve understanding of the catchment processes that drive nutrient fluxes. Furthermore, these data are vital for prediction of future trends under changing environmental conditions and thus the development of appropriate mitigation measures. In recent years, technological developments have led to an increase in the use of in-situ nutrient analysers, which enable measurements at far higher temporal resolutions than can be achieved with discrete sampling and subsequent laboratory analysis. In this paper, we review the principles underlying the key techniques used for in-situ nutrient monitoring and highlight both the advantages, opportunities and challenges associated with high-resolution sampling programs. We then suggest how adaptive monitoring strategies, comprising several different temporal sample frequencies, controlled by one or more ‘trigger variables’ (e.g. river stage, turbidity, or nutrient concentration), can advance our understanding of catchment nutrient dynamics while simultaneously overcoming many of the practical and economic challenges encountered in typical in-situ river nutrient monitoring applications. We present examples of short-term variability in river nutrient dynamics, driven by complex catchment behaviour, which support our case for the development of monitoring systems that can adapt in real-time to rapid changes in environmental conditions. Finally, we suggest future research directions based on emerging technologies in this field.
1. Introduction

Rivers transport and transform nutrients between terrestrial, aquatic, marine and atmospheric systems, thereby playing a key role in global biogeochemical cycling (Ensign and Doyle, 2006; Battin et al., 2008; Hood et al., 2015). Any increase in river nutrient concentrations, whether as a consequence of farming practices or urbanisation, may have substantial implications for aquatic ecosystem structure and functioning (Smith and Schindler, 2009). Eutrophication can result in algal blooms that decrease dissolved oxygen levels, change pH balances, and increase water turbidity. Such changes will impact river habitat and biodiversity, particularly the abundance of sensitive aquatic species (Camargo et al., 2005; Friberg et al., 2010). High river nutrient concentrations may also affect human health and well-being by threatening aquatic ecosystem services, such as freshwater provision for drinking and irrigation, maintenance of fisheries for food, recreational opportunities and aesthetic qualities such as taste, colour or odour (MEA, 2005; Bennett et al., 2009). Nitrate concentrations > 50 mg L⁻¹ in drinking water can cause methemoglobinemia in humans, particularly infants, and may be associated with other adverse health effects including cancer and diabetes (Ward et al., 2005). Consequently, drinking-water treatment plants incur higher costs to reduce excessive nutrient levels to within quality standards, with some organic carbon fractions reacting to form potentially mutagenic and carcinogenic disinfectant by-products during the treatment process (Carpenter et al., 2013).

The environmental impacts of excessive nutrient concentrations highlight the need to understand spatial and temporal variability in river nutrient dynamics. Effective nutrient monitoring strategies can support catchment management by detecting the impact of natural phenomena (e.g. drought) or anthropogenic activities (e.g. land-management practices or point discharges) on river water quality. The resulting information may also help in determining ecological flows and detecting suitable locations or times for water abstraction (Bartram and Rees, 1999; Palmer and Bernhardt, 2006). Long-term continuous datasets of river nutrient concentrations enable the assessment of patterns, trends, and shifts in system behaviour (Burt et al., 2010) and also improve our understanding of the relationships with catchment processes that drive such variability (Bowes et al., 2009). For example, nitrate concentrations measured in both UK and US rivers since the 1930s exhibit marked increases through the post-war period and coincide with increasing applications of inorganic nitrogen fertiliser to arable farmland (McIsaac and Libra, 2003; Burt et al., 2010). More recently, monitoring data have been used to investigate links between river nutrient fluxes and upstream watershed management, demonstrating the potential for land management practices to both elevate and alleviate river nutrient loading (Valiela and Bowen, 2002; Lassaletta et al., 2009).

This information helps any assessment of the effectiveness of mitigation measures to counteract nutrient loading and support future decisions relating to land and river management (Bowes et al., 2009), particularly in the context of predicted future changes in climate that are expected to drive shifts in river nutrient loads (Whitehead et al., 2009). The importance of this is reflected in national legislation and multilateral agreements (e.g. European Union Water Framework Directive or WFD, 2000/60/EC) relating to monitoring, and in some cases also improving, the status of freshwater environments.

River nutrient monitoring has evolved over time from sporadic, ad-hoc, sampling of local rivers (e.g. Casey and Clark, 1979) to the establishment of national river water quality data systems with standardised sampling protocols, such as the UK Harmonised Monitoring Scheme that commenced...
in the mid-1970s (Simpson, 1980) and the General Quality Assessment Scheme that operated from 1990 to 2009 (Environment Agency, 2016) to the implementation of multi-lateral agreements that standardise water quality assessment procedures across member states and, in some cases, across multi-national drainage basins (e.g. European Union WFD; Hering et al., 2010). Traditionally, the frequency and coverage of river water sampling has been constrained by practical issues related to the costs of field personnel and laboratory analysis. Autosamplers enable short-term increases in sampling frequency but have limited volumes and samples may degrade if not preserved or processed quickly (Bende-Michl and Hairsine, 2010). Consequently, the spatial and temporal resolutions of many river nutrient monitoring systems and observational networks have been relatively low, despite a growing body of evidence demonstrating that nutrient concentrations in lotic environments can exhibit highly dynamic and non-linear behaviour in both time and space (Wade et al., 2012; Halliday et al., 2014; Bieroza and Heathwaite, 2015; Krause et al., 2015). Complex nutrient dynamics can arise from variability in climate forcing or changes in environmental management practice as well as accumulation and release of material and associated first flush effects (Krause et al., 2015). For example, increased flows driven by storm events can mobilise sediments and account for a large proportion of the annual total phosphorus export from river catchments (Bowes et al., 2009), while Pellerin et al. (2011) observed high diurnal and event-based variability in the flux of nitrate and dissolved organic matter (DOM) associated with the flushing of shallow hillslope flow paths during spring snowmelt. Such results indicate that river nutrient concentrations are commonly discharge-driven to a large extent, yet often the variability in river hydrological regimes is inadequately captured by the regular frequency of standard water quality sampling programmes. Consequences of inadequate sampling frequencies include missing key pulses of nutrients during short-term events, and potentially underestimating long-term nutrient fluxes by up to an order-of-magnitude (Mellander et al., 2012; Neal et al., 2012; Krause et al., 2014).

In recent years, developments in sensing technologies, coupled with decreasing costs, have led to increased usage of in-situ analysers to monitor key macronutrient (i.e. C, N, P) concentrations, species (e.g. NO$_3$, NO$_2$, NH$_4$) and composition (e.g. DOM; spectral slope and fluorescence spectra) (Figure 1). This technology enables monitoring at far higher frequencies than can be achieved with discrete sampling and subsequent laboratory analysis and, importantly, allows the pairing of nutrient and river flow measurements at the same temporal resolution. Data generated from in-situ high-resolution (i.e. frequencies of 10$^{-1}$ to 10$^{-3}$ h) monitoring (HRM) can yield valuable new insights into the drivers, controls and organisational principles of nutrient dynamics in lotic ecosystems (Bowes et al., 2009), and may help to identify future responses to environmental pressures and potential tipping points in system behaviour (Wade et al., 2012). In addition to characterising nutrient fluxes at catchment outlets, deconvolution of HRM data may also enhance our capacity to discover more about the time-variant activation of catchment source area dynamics (Krause et al., 2014). HRM also has potential applications in improving legislative monitoring strategies (Halliday et al., 2015), enhancing water body classification schemes (Skeffington et al., 2015) and providing an early warning of potential harmful by-products or bacteria (Bridgeman et al., 2015; Li et al., 2016). However, HRM equipment can be costly to run and difficult to maintain, and HRM is not necessarily appropriate for all river nutrient monitoring applications. This paper aims to outline the principles of measurement and the major advantages and disadvantages associated with contemporary in-situ nutrient sensing technologies, and demonstrate the potential for real-time adaptive monitoring to provide new insights into processes operating within river catchments whilst overcoming some of
the challenges associated with the collection and analysis of data generated through HRM. We use
case studies to demonstrate the applicability of this approach to a diversity of environments. Finally,
we conclude by highlighting key areas for future research where *in-situ* monitoring data may
improve understanding of nutrient dynamics and ecosystem processes within river catchments.

**Figure 1**: Number of articles published by year listed on Web of Science matching the following
search terms: (high-resolution OR continuous OR automated) AND (nutrient OR NO$_3$* OR nitrate OR nitrogen OR
ammonia OR ammonium OR NH$_3$* OR NH$_4$* OR doc OR *dom OR tryptophan OR PO$_4$* OR phosphate OR phosphorus) AND
(sampling OR monitoring OR sensor OR analy*) AND (stream OR river). Data from 2016 are excluded to avoid
potential issues with as-yet-uncatalogued articles.
2. Principles of in-situ nutrient monitoring techniques

In-situ determinations of macronutrient concentrations, species and compositions utilise recent technological advances in electrochemical detection, colorimetry and optical methods using absorbance or fluorescence, which offer increasing opportunities for HRM. Table 1 summarises the parameters which are generally measured via each technique, along with the measurement ranges and the associated accuracy.

<table>
<thead>
<tr>
<th>Table 1: Summary of commonly-used measurements methods for in-situ nutrient monitoring and the associated accuracy, range and maintenance requirements for each method</th>
</tr>
</thead>
</table>

2.1 Electrochemical detection

The use of electrochemical detection for nutrient determination using Ion-Selective Electrodes (ISEs) is well established and the technology has been miniaturised to enable in-situ quantification of NO$_3^-$, NH$_3$, and pH (Le Goff et al., 2002; Merks, 1975; Collings, 2003). ISEs use direct potentiometry between a sensing electrode and a reference electrode (Figure 2a) with measurement ranges typically spanning at least three orders of magnitude (YSI, 2016; Hach, 2016). Unlike many optical techniques (see below), ISEs are not influenced by water colour or turbidity, but are often subject to significant drift and ionic interferences (de Marco et al., 2007) and can also suffer from systematic errors if the ionic activity of the calibration buffer solution is markedly different to that of the field sample (Le Goff et al., 2002). However, newer sensors have been developed which are able to combat some of the biggest problems with ISEs. For example, Le Goff et al. (2002) describes a nitrate ISE that measures over a reduced concentration range but which remains stable for up to four months in the field, and Tang et al. (2012) present a solid-state ISE for analysis of NO$_3^-$.

2.2 Colorimetry

Colorimetric methods have been developed to quantify NO$_3^-$, NO$_2^-$, NH$_4^+$, PO$_4^{3-}$ and total phosphorus (TP) using either submersible microfluidic technology or bank-side analysers. Samples are mixed with reagents to generate coloured solutions which are then detected using photometry (Figure 2b). Colorimetric techniques generally have higher levels of precision and accuracy than ISEs, however their measurement range is reduced and usually spans one to two orders of magnitude (Patton and Kryskalla, 2011). Importantly, most colorimetric reactions are temperature-dependent and post-correction is required if water temperatures change significantly during deployment. In addition, sampling resolutions are constrained by the reaction time necessary for the colour change to completely develop, and turbid waters can influence measurements by altering the background water colour.

2.3 Optical UV-VIS spectroscopy
Optical sensors using ultraviolet spectroscopy provide a reagent-free method for determining a range of parameters including dissolved organic carbon (DOC) and NO$_3$. The measurement principle is based on the spectral absorption of a sample at a defined wavelength (Figure 2c). Appropriate algorithms are necessary to convert the optical signal to concentration of the parameter of interest, and hence it is a proxy technique rather than a direct measurement of the parameter. The resulting data are usually of high resolution and highly precise and accurate, although this can vary depending upon the parameter required (van den Broeke et al., 2006). Potential problems with optical interferences are similar to those associated with colorimetry when deployed in river water due to high sediment loads, debris and discoloration of the water. However, many commercially-available devices now incorporate algorithms to correct for interference by using secondary compensation wavelengths or, in some cases, the whole UV-VIS absorption spectrum.

2.4 Optical fluorescence spectroscopy

Optical sensing using fluorescence has grown in popularity due to technological advances and reduced in-situ sensor costs. Single excitation-emission sensors quantify the light emitted from a sample at a particular wavelength when the sample is excited by a known wavelength of light (Figure 2d). While the in-situ sensors have a narrow wavelength range compared with their benchtop equivalents, they can target particular compound classes based on their fluorescent properties. Typically sensors are used to detect relatively low molecular weight compounds such as proteineous material and hydrocarbons, but can also be used to identify more complex macromolecular material which makes up the coloured DOM (CDOM) fraction. Specific wavelengths can be used to quantify parameters such as chlorophyll $\alpha$ and dissolved oxygen (DO). The measurements are rapid and have the potential to yield sub-five minute resolution data. The use of fluorescence has similar issues to UV optical sensors and is subject to interferences from turbidity (Saraceno et al., 2009), bubbles, temperature (Watras et al., 2011), shading and fouling. It is also important to determine absorbance at the relevant wavelengths to enable data post-processing as inner-filter effects caused by attenuation of excitation light and reabsorption of emitted photons (Downing et al., 2012; Khamis et al., 2015; Lakowicz, 2013).
Figure 2: Diagrammatic representation of principles of in-situ nutrient monitoring techniques for a) electrochemical, b) colorimetry, c) optical UV-VIS spectroscopy, and d) optical fluorescence spectroscopy.

2.5 Practical considerations

HRM is a powerful tool for hydrological and water quality research, although maintenance frequency is proportional to the sampling frequency and hence HRM may require more frequent maintenance compared with lower resolution sampling campaigns. To ensure the highest quality data from HRM systems, several issues must be considered to determine the most appropriate sensor type and monitoring location.

Firstly, power demand varies between sensor types, dataloggers and telemetry systems. Power is usually provided through batteries, fuel cells or solar panels, although for some bankside analysers mains power is the most suitable option given the higher power requirements associated with pumping water to the analyser. For batteries or fuel cells, regular checks are necessary to minimise instrument drop-outs and changes may be required once or more each month, particularly during winter when batteries become less efficient at lower operating temperatures. Solar panels provide a more sustainable alternative, although suitable siting is important as shaded areas may yield insufficient power generation, especially in winter when solar altitudes are low. Second, virtually all in-situ sensors are subject to fouling. Most in-situ sensors are submerged in the water body and can be susceptible to accumulation of sediment, biofilms, or other debris around the sensor. Fouling can also affect bankside setups, particularly in the tubing which carries the sample to the sensor. Technologies to reduce the impact of biofouling on sensors are now commercially available,
including anti-fouling coatings, automatic wipers or compressed air which seek to prevent, or clear, accumulated particulates and biofilm growth (Kotamäki, 2009; Conmy et al., 2014). However, regular manual cleaning is required for most monitoring systems and should be considered a mandatory component of HRM programs (Jones et al., 2014). Third, most sensors are affected to a degree by other environmental factors such as pH, temperature, electrical conductivity and turbidity (e.g. Downing et al., 2012; Khamis et al., 2015). These environmental variables vary through time and, unless explicitly accounted for by the sensor, should be monitored alongside the parameter of interest so that post-processing corrections can be applied. Furthermore, although routine onsite calibrations can achieve increasingly accurate measurements (Saraceno et al., 2009), sensor outputs should be ‘ground-truthed’ using samples that are independently measured under controlled conditions in the laboratory.

Careful consideration of these issues should minimise sensor down-time and ensure accurate and high-quality data output. Inevitably, however, sensors will be subject to occasional mechanical failure, particularly when deployed to monitor nutrient behaviour during storms where the risk of damage is increased. Therefore, sensor checking should be carried out routinely so that problems can be quickly identified and fixed. Telemetry systems are invaluable in this respect as sensor performance or power supply issues can be quickly identified (Glasgow et al., 2004).

3. High resolution in-situ river nutrient monitoring

3.1 Opportunities and challenges

The use of in-situ sensing technologies to monitor nutrients and dissolved organic matter (DOM) in riverine environments has lagged behind other aquatic systems (Johnson et al., 2007; Saraceno et al., 2009). In-situ monitoring of absorbance and fluorescence to investigate the spatial and temporal dynamics of DOM, nitrate and primary production were first undertaken ~20 years ago in marine systems (Daniel et al., 1995; Pega et al., 1995; Chen, 1999). This early work typically involved relatively large instruments with high power consumption, and their deployment was largely restricted to research vessels (e.g. Daniel et al., 1995). Much of this instrumentation was engineered to withstand high pressures to enable deep sea deployment (e.g. Finch et al., 1998) and the increased cost limited applications of this technology to marine systems. However, recent advances and decreased costs have facilitated the use of HRM equipment in freshwater systems to improve understanding of nutrient and DOM dynamics for a range of freshwater ecosystems, including urban (Wade et al., 2012; Halliday et al., 2015), agricultural (Bieroza et al., 2014; Outram et al., 2014) and alpine rivers (Pellerin et al., 2011). Advances in field deployable monitoring technologies are enhancing our ability to understand the linkages between climate, catchment properties, and hydrological/ biogeochemical fluxes (Krause et al., 2015) and to detect environmental change at finer spatial and temporal scales (Sobczak and Raymond, 2015). This section will outline the capacity and limitations of HRM in the context of improving hydro-ecological understanding of lotic systems and associated interfaces.
The ‘traditional’ approach for monitoring nutrient and pollutant loads has involved the collection of discrete spot samples, either manually or via autosamplers, to estimate fluxes and annual loads (e.g. Johnes, 2007). However, while monthly or fortnightly sampling regimes may be sufficient to characterize lentic systems (Pobel et al., 2011), they are unable to adequately represent the dynamic behaviour of lotic system potentially contributing to serious errors in load estimation (Johnes, 2007; Cassidy and Jordan, 2011; Lloyd et al., 2016). For example, while weekly spot sampling may be sufficient to detect inter-annual variability in N and P loads, significant information on the flow extremes (i.e. shape of the frequency distribution) may be lost when compared to sub-hourly monitoring (Bieroza et al., 2014). Other studies highlight the need to consider catchment behaviour (i.e. flashy vs muted flow response to storm events) when determining monitoring frequency; for example, Cassidy and Jordan (2011) suggested that for a flashy catchment sub-hourly (i.e. 20 mins) HRM is required to accurately quantify the P load as even storm event triggered sampling led to high uncertainty. However, Lloyd et al. (2016) highlighted the need for a systemic approach to handling errors associated with HRM flux estimates (N and P) as, in many cases, sampling at high resolution results in tradeoffs between measurement precision and accuracy.

Significant pulses of nutrients and DOM can be rapidly mobilized and transported through the river network during storm events (Hinton et al., 1997). Recently, in-situ HRM using field deployable UV-absorbance sensors has provided new insight into C dynamics during storm events (e.g. Jeong et al., 2012; Strohmeier et al., 2013; Table 2). Rainfall intensity has been identified as a key control on the form of C transported (i.e. dissolved vs particulate) and HRM has enabled C sources and flow paths to be traced (Jeong et al., 2012) thus improving our understanding of how these systems are likely to respond to climatic change. Furthermore, recent sub-hourly monitoring of N & P in the UK has highlighted important links between meteorological conditions (event and antecedent conditions) and in-channel nutrient fluxes (Outram et al., 2014; Table 2). Such information is needed when developing adaptation strategies for nutrient enriched catchments, particularly given anticipated changes in the magnitude and frequency of extreme meteorological and hydrological events (Kendon et al., 2014).

In addition to event-based nutrient dynamics, HRM can also offer new insights into hydro-ecological processes at temporal scales ranging from diurnal to seasonal. For example, in-situ fluorometers have enabled identification of distinct diurnal dynamics of organic C in rivers (Spencer et al., 2007; Worrall et al., 2015; Table 2) and lakes (Watras et al., 2015). Biological activity was suggested as the primary driver of these patterns (Spencer et al., 2007; Watras et al., 2015); however, Spencer et al., (2007) suggest that photolysis may also be an important mechanism. Moreover, Worrall et al., (2015) focused on the links between abiotic factors and diurnal DOC cycles for the River Dee, UK. Using continuous UV absorbance data they suggested residence time and the accumulated temperature of water parcels (linked to day length) were the main drivers of diurnal patterns. In-situ FDOM monitoring (ex. 365 nm; em. 470 nm) coupled with discrete sampling and laboratory analysis revealed distinct seasonal changes in DOM composition (i.e. higher aromatic/molecular weight during the freshet period) for a river draining a mountainous river basin (Voss et al., 2015). Wilson et al. (2013) also employed in-situ FDOM sensors to reveal seasonal changes in event based DOC fluxes. Distinct peaks in DOC quantity and bio-availability were associated with summer storm events highlighting the utility and advantage of in-situ HRM for tracking long term changes in DOC flux.
Large scale monitoring projects/schemes for legislative purposes (e.g. for the WFD) were devised based on data obtained through discrete sampling and subsequent laboratory analysis. Through the increased availability of HRM data it is becoming increasingly apparent that the limited sensitivity of traditional sampling methods leads to significant errors in WFD chemical class assignment, constraining our ability to detect trends (Skeffington et al., 2015). For example, HRM of NO₃ and TP on the River Wylye (Wiltshire, UK) showed that annual loads can be significantly underestimated if only daily sampling regimes are used (Lloyd et al., 2016). High frequency monitoring of an urban stream in southern England identified distinct diurnal (in stream production) and seasonal (growing season) variability in parameters used to classify WFD chemical status (e.g. temperature, dissolved oxygen and P). This highlights the need to specify specific windows for WFD sampling as spot sampling in dynamic river systems can lead to spurious inferences regarding WFD status (Halliday et al., 2015). Bowes et al. (2012) used high resolution P records to differentiate between sewage treatment works (STW) and diffuse and in-channel sources to confirm the STW was operating within consent limits. This research suggests that in addition to increasing the resolution of routine legislative monitoring in some cases, the way that WFD limit criteria are designed should also be reviewed where and when necessary. For example, WFD limits for soluble reactive phosphorus (SRP) are currently based around mean annual concentrations (UKTAG, 2013), the calculation of which is highly impacted by the sampling strategy. Therefore, there is an argument for developing more intelligent and robust limit guidelines for classifying water quality status. Furthermore, there is still a lack of a WFD classification in the UK for NO₃ and at present the Drinking Water Directive (98/83/EC) acts as a guideline for river water NO₃ concentrations but does not explicitly define limits required to meet good status.

There is increasing evidence that HRM data will improve our understanding of catchment nutrient dynamics particularly during ‘active’ periods. For example, simple hysteresis indices have been revised to improve understanding of nutrient transport processes in agricultural catchments (Bieroza and Heathwaite, 2015; Lloyd et al., 2016). In addition, contemporary sensors provide the means to identify and monitor DOM processing in-situ, particularly as advances in LED technology mean monitoring using a suite of in-situ optical techniques (i.e. fluorescence and absorbance at different wavelengths) is becoming economically viable. The more detailed optical map provided can offer new insight into DOM dynamics in riverine environments enabling us to monitor both DOM quality and quantity continuously (Spencer et al., 2007). Yet there are still significant challenges in achieving accurate in-situ measurement of DOM and other nutrients and these new UV-sensors should be seen as complementing, not replacing, traditional grab sampling (Sobczak and Raymond, 2015). There is a particular need to compensate for temperature and turbidity interference (Downing et al., 2012; Khamis et al., 2015) and account for seasonal variability in background water composition (Wilson et al., 2013). Subsequent analysis of these high resolution data also requires careful consideration as the assumptions of many statistical tests are violated for time series data due to temporal auto-correlation and heteroscedasticity (Lloyd et al., 2014). Furthermore, the maintenance and calibration schedules need to be carefully considered to ensure data quality as, even in pristine systems, fouling can lead to sensor drift over relatively short time intervals (Jones et al., 2014). However, in some cases in-situ analysis can yield more accurate measurements than those produced by more traditional analytical methods. For example, Bieroza and Heathwaite (2016) demonstrated that measurement errors arising from short-term chemical changes in grab samples (i.e. those that
occur between sample acquisition and laboratory analysis) masked diurnal patterns in P and OM that were otherwise detected by in-situ analysers.

Table 2: Summary of published studies on in-situ nutrient monitoring highlighting study duration, nutrients investigated, analysis methods employed and time required for processing

3.2 Adaptive monitoring: a way forward?

While current studies using HRM are providing new insights into nutrient/DOM dynamics at a range of spatiotemporal scales, monitoring resolution is often limited by power consumption and data storage capacity (Downing et al., 2009). Given the pulsed behaviour exhibited by many aquatic ecosystems (Junk et al., 1989; Tockner et al., 2000) conventional modern HRM schemes operating at hourly or sub-hourly resolution can result in significant data redundancy. For example extended periods of low biogeochemical activity or variability often occur during low flow periods (see Figures 3a and 3b). The unnecessarily high resolution of sampling can result in excessive memory and power consumption, and for certain monitoring techniques can increase maintenance requirements and waste generation (i.e. P analysers; Table 2). Conversely, during biogeochemically ‘active’ periods (e.g. high flow events) the resolution of many conventional HRM schemes is too coarse for detailed event characterization (Figure 3b). Thus, for monitoring dynamic riverine systems or examining eco-hydrological processes that exhibit strong non-linear properties, an adaptive monitoring approach is required (cf. Krause et al., 2015). Here differing monitoring frequencies can be triggered based on the variable of interest (e.g. N or P), a surrogate/driver variable (e.g. level, electrical conductivity, turbidity, rainfall) or combination of variables. For example two thresholds (TT1 & TT2) can be defined based on the concentration of a nutrient of interest (Figure 3c, 3d, 3e). Using these thresholds as triggers to alter the monitoring frequency enables the capture of (i) the first flush and peak flow at very high resolution, and (ii) the recession at moderate resolution. If necessary, an additional threshold (TTa), such as river level, can be included to avoid triggering a change in monitoring frequency due to anomalous sensor readings. In this scenario the nutrient concentration (TT1) and river stage (TTa) must both be exceeded to initiate a change in monitoring frequency (Figure 3a and 3c). Furthermore, to avoid a ‘yo-yo’ effect, where the monitoring state alternates rapidly between two frequencies, additional logic commands can be included (e.g. switch from state TT2 to TT1 only when the previous three readings fall below a defined threshold). Finally, as telemetry has become more versatile and reliable, monitoring frequencies can reliably be changed remotely and in real-time, and warning alarms can also be triggered if defined thresholds are breached (Othman and Shazali, 2012).

The science questions to be addressed are particularly important when setting thresholds for adaptive monitoring. Non-linearity and ‘hot-moments’ may be associated with different atmospheric, management, or accidental drivers (Krause et al., 2015). For example when exploring atmospheric drivers such as precipitation, the river level or discharge would be an appropriate trigger variable given the focus on storm event dynamics. However, accidental releases (e.g. sewer
discharge, river bank collapse) would require a different trigger variable. While the parameter of interest could be used (see Figure 3d), the short windows associated with accidental release (Carstea et al., 2010) could lead to an event being missed if it occurred between the coarse monitoring time steps (Figure 3c). In this case a surrogate variable with lower maintenance requirements could be monitored at high frequency (e.g. specific conductance or turbidity) providing a trigger for the nutrient analyser. Knowledge of the system is also particularly important for an effective adaptive monitoring strategy as catchment area and land use are both important and will influence the lag time between trigger and response (see section 4).

Figure 3: Conceptual representation of adaptive monitoring principals based on Krause et al (2015). Vertical dashed lines represent sample points, TT1 is trigger threshold 1 (black line), TT2 is trigger threshold 2 (red line) and TTa is an additional trigger threshold (horizontal dashed line). a) Hydrological context (precipitation and river level) and TTa, b) ‘traditional’ high resolution monitoring of a variable (nutrient) of interest, c) adaptive monitoring approach with TT1 and TT2
highlighted, (d & e) events of interest or ‘active periods’ highlighting the transition between the	hree monitoring frequency states based on threshold exceedance under rapidly changing
environmental conditions. Note for a change from baseline monitoring frequency to higher
resolution monitoring states TT1 & TTa must be exceeded to avoid inappropriate triggering.
4. Catchment variability in nutrient dynamics

In this section we present examples showing how novel monitoring strategies have delivered significant advances in process understanding and highlighted the variability in river nutrient dynamics in response to short-term drivers of change. In so doing, our aim is to demonstrate the extent and diversity of catchment differences in nutrient responses to which HRM could potentially be applied, thereby emphasising the need to develop site-specific sampling strategies (including thresholds and trigger variables) at frequencies appropriate to the parameters and system dynamics of interest. These thresholds should be reviewed regularly and updated where necessary to account for system behaviour changes caused by environmental, climate, or anthropogenic drivers. The examples illustrate the potential for adaptive real-time HRM strategies to capture nutrient dynamics during key hydrological events and hence improve our understanding of the catchment processes that underpin these patterns.

4.1 River Wear, Durham, UK

The River Wear, Northern England (length = 106.9 km; catchment area: 1008 km²) drains an area of the Eastern Pennines (see Neal et al. (2000) for a detailed description). During the autumn of 2015 CDOM fluorescence was monitored at the raw water intake of a water treatment works, close to a gauging station at Chester le Street, Northumbria (54°51’N, 1°33’W). The catchment is predominately grassland/moorland (~60 %) with minimal urban cover (<5 %) and a median altitude of 213 m. During the study period a mean lag time of ~18 hrs between rainfall and peak flow was observed (Figure 4). A significant pulse of coloured dissolved organic matter (CDOM) was recorded after rainfall events between 23/09/15 and 24/09/15. However, peak CDOM lagged peak discharge by 56 hr, suggesting the primary source of CDOM at this site is the organic rich soils and peats in the upper catchment (Neal et al., 2000). Thus, the delayed peak is likely due to a combination of: (i) wetting up time (i.e. time taken for shallow flow paths to activate); and, (ii) transit time to the monitoring site ~60 km from the headwater CDOM sources (Hangen et al., 2001; Inamdar and Mitchell, 2006). In this case, due to the apparent disconnection between river flow and CDOM concentration, precipitation and discharge would not be suitable triggers for changing monitoring frequency (adaptive HRM). However, due to the lengthy rising and falling limbs of the CDOM chemograph, the variable of interest would be an effective trigger. Here, given the lagged and relatively gradual response, it would be possible to represent the CDOM chemograph variability with relatively low frequency sampling (i.e. hourly).
Figure 4: High resolution (15 min), in-situ, fluorescence and hydrological monitoring records from the River Wear. CDOM (coloured organic matter) was measured using a submersible CDOM fluorometer (Cyclops 7, Turner Inc.; Ex. 365nm, Em.460nm). A strong relationship between Total Organic Carbon (TOC) and CDOM was recorded ($R^2 = 0.97$, $n=11$). An example of adaptive monitoring is shown by grey bars that denote a switch from low to high sampling frequency triggered by CDOM exceeding a defined threshold ($T1$).
4.2 Bourn Brook, Birmingham, UK

The Bourn Brook, a headwater tributary of the River Rea, is a small lowland stream (length = 7.4 km) in central England. During the autumn of 2014 a monitoring station recording river level and tryptophan-like fluorescence (TLF) was established close to the University of Birmingham campus (52°27′N, 1°54′W). The upstream catchment of 27.9 km$^2$ is mainly urban with a median altitude of 169 m asl. The site displays a distinct urban hydrology (Gurnell et al., 2007) with a flashy flow regime and <2h lag time to peak flow (Figure 5). The organic matter dynamics for this site display a classic first flush effect (Lee and Bang, 2000), with TLF peaking either on the rising limb or at peak flow (Figure 5b). This suggests DOM sources are situated close to or in the channel, and probably include dry deposition on impervious surfaces and the breakup of biofilms in the channel, storm drains and combined sewer overflows (Carstea et al., 2009; Khamis et al., 2015). Thus, for this system, suitable triggers for adaptive monitoring would be a combination of river level/discharge and TLF (cf. Fig. 3). However, it is important to consider the purpose of the investigation as the adaptive monitoring approach outlined above would miss point source pollution event dynamics because the Boolean argument (level AND TLF) would not be satisfied.
Figure 5: High resolution (15 min), in-situ, fluorescence and hydrological monitoring records from the Bourn Brook. TLF (tryptophan-like fluorescence) was measured using a Cyclops 7 fluorometer (Ex. 285 nm, Em. 350 nm). A strong relationship between Biochemical Oxygen Demand (BOD) and TLF was observed ($R^2=0.92$, n=21). An example of adaptive monitoring is shown by grey bars that denote a switch in sampling frequencies controlled by Boolean logic as follows: level $> T_1 =$ high frequency monitoring (light grey bars); level $< T_1$ AND TLF $> T_2 =$ medium frequency monitoring (dark grey bars); level $< T_1$ AND TLF $< T_2 =$ low frequency monitoring.
The Mill Brook, central England, drains 3.5 km² of arable farmland and a mixture of mature and new-growth deciduous woodland. The stream flows through the Birmingham Institute for Forest Research (BIFoR) field site where a permanent monitoring station was established in January 2015. A suite of water quality variables are currently analysed in-situ at 10-minute resolution, including spectrophotometer measurements of NO₃-N and fluorometer measurements of CDOM and TLF. Data recorded to date suggest the system responds rapidly to precipitation events and that nutrient dynamics are driven strongly by changes in discharge (Figure 6). Both NO₃-N and CDOM decrease markedly in response to increased discharge, whilst TLF increases with discharge. However, HRM allows shorter-term nutrient dynamics to be identified, most notably an initial increase in NO₃-N concentration associated with the onset of precipitation events. This may be driven by the mobilisation and flushing of nitrate-rich waters in agricultural tile drains, which are common in agricultural watersheds (Wagner et al., 2008). At Mill Brook, an increase in monitoring frequency (e.g. from hourly to 10 minute) triggered either by precipitation or increasing stream stage is necessary to capture the role of specific hydrological events for nutrient export (particularly NO₃) from the catchment.
Figure 6: High-resolution 10 minute monitoring data from the Mill Brook at the Birmingham Institute for Forest Research (BIFoR) field site in Staffordshire, UK. NO$_3$-N is determined using an optical Trios Opus UV spectrophotometer. CDOM (chromophoric dissolved organic matter) and TLF (tryptophan-like fluorescence) are measured using Turner Inc. Cyclops-7 fluorometers (Ex. 325nm, Em. 460nm and Ex. 285 nm, Em. 350 nm, respectively). An example of adaptive monitoring is shown by grey bars that denote a switch from low to high sampling frequency initiated when precipitation > T1 and ended when level < T2.
4.4 River Wylye, Wiltshire, UK

The River Wylye, a chalk stream in Southern England, is a headwater tributary of the Hampshire Avon. A monitoring station was installed at Brixton Deverill in 2011 as part of the Demonstration Test Catchment Programme, which aimed to use HRM of hydrochemistry to test the efficacy of on-farm pollution mitigation strategies (McGonigle et al. 2014). The station was located close to an existing gauging station (EA gauge 43806) which provided 15 minute resolution discharge measurements. At this point, the river drains 50 km² of agricultural land dominated by intensive mixed arable farming. The catchment hydrology has a significant groundwater contribution, resulting in slow baseflow variations in response to rainfall. Superimposed on this signal are flashy hydrological responses arising from direct input of precipitation and local surface runoff. The storm events illustrated in Figure 7 show a rapid response in both NO₃-N and TP to increasing discharge. NO₃-N shows a dilution response due to surface-water input, as the main source of NO₃-N in the river is groundwater. Conversely, storm events resulted in pulses of TP at the monitoring station. HRM allowed the change in the lag between peak discharge and peak TP to be monitored across storms. Of the four storms shown here, the first storm resulted in an immediate TP response, most likely due to the remobilisation of bed sediments deposited at lower flow (Ballantine et al. 2009; Dorioz et al. 1998). Subsequent storms show an increasing lag between the discharge peak and the TP peak indicating a longer transit time for the pulse of TP to reach the monitoring station (Lloyd et al., 2016; Bowes et al. 2005). These data highlight the importance of HRM to understand the transport mechanisms responsible for nutrient delivery to the stream during storm events. This also supports the argument for adaptive sampling strategies so that intense monitoring is undertaken only when it provides the maximum information about system processes driven predominantly by short-term event-based dynamics.
Figure 7: High-resolution 30-minute monitoring data from the River Wyle at Brixton Deverill, collected as part of the Demonstration Test Catchment Programme. Discharge data were provided by the Environment Agency (England) and other data were provided by the Hants Avon DTC consortium (2016). \( \text{NO}_3^-\text{-N} \) was determined using an optical UV Hach Lange Nitratax sensor and TP was measured using a Hach Lange Phosphax Sigma bankside analyser which uses colorimetry following sample digestion. An example of adaptive monitoring is shown by grey bars that denote a switch from low to high sampling frequency triggered by TP exceeding a defined threshold (T1).

5. Conclusions and future directions

Recent technological advances in a number of areas have significantly enhanced our ability to monitor the major nutrients and DOM in rivers in real-time. This has the potential to revolutionise our capacity to understand and quantify the complex spatial patterns and temporal dynamics of biogeochemical cycling in systems sensitive to environmental change. Such developments facilitate insights into the wider significance of nutrient cycling and transformation processes, in particular at
biogeochemical hotspots and during hot moments of biogeochemical activity. Extending our perspective to fully capture the dynamic behaviour of catchment nutrient fluxes and transformations requires wider, integrated, application of the sensing technologies outlined above. For example, further development of semiautonomous sensing networks providing data at sufficient spatial and temporal resolution to monitor and model catchment biogeochemistry and hydrology and their links with ecosystem functions represent important advances (Crawford et al., 2015). In particular, improvements in telemetry and developments in adaptive sampling have the potential to provide a step change in our understanding of dynamic systems but the potential of the currently available technologies has yet to be fully utilised. The examples discussed in this review illustrate some of the advances in process understanding that are possible by integrating new technologies for determining nutrient concentrations in aquatic systems with high resolution monitoring systems. The ability to specify particular triggers for sample collection, or for changing the temporal resolution of data collected, has the potential to make autonomous in-situ sampling more efficient and enhance our ability to capture system behaviour: quantifying the importance of fluxes associated with extreme hydrological events and yielding improved estimates and understanding of nutrient cycling through river catchments from their headwaters to the freshwater-marine transitional zone and beyond.

Improving our understanding of complex nutrient cycles and their interactions across ecosystem boundaries represents significant interdisciplinary challenges. For example, whilst HRM can be invaluable in improving our estimates of nutrient fluxes, there remains a need for greater cross-disciplinarity across the different facets of aquatic monitoring programmes. Historically there has been a distinct mismatch between standard ecological sampling protocols and nutrient/chemical monitoring schemes. As a result, potentially important feedbacks between ecology and biogeochemical cycling may be missed. However, promising advances in real-time, automated species recognition tools (e.g. Aide et al., 2013) could herald a new era of ecosystem monitoring in which our ability to monitor physiology and movement could be complemented by real-time nutrient monitoring (Kuklina et al., 2013).

To a considerable extent, our understanding of these processes to-date has been constrained by the distribution and density of monitoring locations, which in many cases reflect high instrumentation costs and limited funding availability. Recent and future developments of low cost and open-source sensors for water quality monitoring have enormous potential here; particularly the combination of micro-controllers and 3-D printers. For example, Leeuw et al., (2013) outline the development of an in-situ fluorescence sensor for chlorophyll $a$, with a unit cost (in 2013) of US$ 150, while Wittbrodt et al., (2015) provide details of a low cost nitrate sensor using the Nitrate Reductase Nitrate–Nitrogen Analysis Method. Given the significantly reduced cost of these new open source products, greater densities of in-situ riverine nutrient monitoring networks should be feasible, matching advances in urban air quality monitoring (Mead et al., 2013).

Further exciting advances can be expected by fully utilising the potential to link outputs from autonomous sensor networks to Earth Observation (EO) platforms. Recent and ongoing developments in EO provide opportunity for enhanced water quality monitoring and assessment in freshwaters, transitional zones, through to marine environments (Tyler et al. 2016). While EO data are now used routinely for marine observation, river applications are constrained by spatial resolution, and costs for data acquisition, particularly for new platforms (e.g. Dugdale et al., 2015).
However, the spatial resolution of new sensors and platforms, such as PACE / Worldview-3, at >1m, opens opportunities to capture spatial trends in nutrient dynamics (e.g. CDOM) or proxy indicators, such as turbidity as a proxy for TP (Viviano et al., 2014). Inevitably, however, these applications will rely upon extensive field verification with appropriate quality assurance and control. This may be facilitated by automated sensor recalibration, but given the spatial and temporal variability in nutrient transfer, catchment-specific calibrations of individual sensors may be necessary (cf. Khamis et al. 2015) and time-invariance in sensor calibration parameters should not be assumed.

As data of increasing spatial and temporal resolution become available, the maintenance of long-running data-sets is becoming increasingly important (Holmes, 2006; Burt et al., 2010) to capture the extent of ‘natural’ environmental variability (Crawford et al. 2014) and to develop hypothesis-driven science and monitoring to advance our understanding of system behaviour. There is now scope for the wider application of recent technological advances to provide greater insight into the wider biogeochemical functioning of river catchments, particularly with reductions in sensor costs, and increasing availability of EO data. If practical problems, for example with respect to power supply and biofouling, can be resolved, these developments offer a potential paradigm-shift in our understanding of environmental complexity (cf. Krause et al. 2015), our ability to characterise the fluxes of key nutrients from catchments-to-coast, and our understanding of the spatial variability in freshwater biogeochemistry and hydrology.

Acknowledgements

We thank the Birmingham Institute of Forest Research (BIFoR), the Environment Agency (England) and the Hants Avon DTC consortium for contributing datasets. We are also grateful to three anonymous reviewers who provided helpful comments on an earlier version of the manuscript.
References


of DOM fluorescence in rivers and streams. *Limnology and Oceanography: Methods*, 10(10), 767-775.


Watras, C., Morrison, K., Crawford, J., McDonald, C., Oliver, S., & Hanson, P. (2015). Diel cycles in the
fluorescence of dissolved organic matter in dystrophic Wisconsin seepage lakes: Implications
for carbon turnover. *Limnology and Oceanography*, 60(2), 482-496.

impacts of climate change on surface water quality. *Hydrological Sciences Journal*, 54(1),
101-123.

Wilson, H. F., Saiers, J. E., Raymond, P. A., & Sobczak, W. V. (2013). Hydrologic drivers and
seasonality of dissolved organic carbon concentration, nitrogen content, bioavailability, and

Photometric System for Enzymatic Nitrate Quantification. *PloS one*, 10(8), e0134989.

carbon–The interplay of in-stream residence time, day length and organic matter turnover.
*Journal of Hydrology*, 523, 830-838.

Electrodes-W74-1014-spec-sheet.pdf
<table>
<thead>
<tr>
<th>Measurement method</th>
<th>Measurement principal</th>
<th>Parameters</th>
<th>Measurement range</th>
<th>Accuracy</th>
<th>Maintenance</th>
<th>References</th>
</tr>
</thead>
</table>
| Absorbance          | Spectrophotometric method to measure the amount of light of a specific wavelength absorbed by a sample | Nitrate/nitrite | 0.1-500 mg L⁻¹ NO₃-N | Up to 10% of reading | • Regular calibration (3-6 months)  
• Regular cleaning due to fouling  
• Check power supply  
• Check light source | Huebsch et al., 2015 |
|                     |                        | TOC/DOC    | 1-150 mg L⁻¹ TOC 0.5-75 mg L⁻¹ DOC | <10% of reading |                        | Waterloo et al., 2006; Jeong et al., 2012; Lee et al., 2015 |
|                     |                        | Phosphate  | 0-0.3 mg L⁻¹ PO₄-P | 2% of reading |                        | Subchem |
| Fluorescence        | Spectrophotometric method to measure the emission of light at specific wavelengths after excitation with a specific wavelength of light. | Tryptophan-like | 0-20000 ppb | 2% of reading | • Regular calibration (once a month)  
• Regular cleaning due to fouling  
• Check power supply  
• Check light source | Khamis et al., 2015 |
|                     |                        | CDOM       | 0-500 µg L⁻¹ | ± 0.09 µg L⁻¹ |                        | Chen, 1999 |
|                     |                        | Chlorophylla | 0-125 µg L⁻¹ | ± 0.02 µg L⁻¹ |                        | Del Castillo et al., 1999 |
| Colormetric         | Wet chemical technique, detection via photometry | Nitrate/nitrite | 0.3-50 mg L⁻¹ | <3% of reading | • Regular calibration (once a week or once a day if automated)  
• Regular cleaning due to fouling  
• Check power supply  
• Replacement of reagents  
• Disposal of reagents | Greenway et al., 1999; Petsul et al., 2001 |
<p>|                     |                        | Phosphate  | 1-10 mg L⁻¹ | &lt;5% of reading |                        | Doku and Haswell, 1999, Cleary et al., 2008 |
|                     |                        | Total phosphorus | 0.1-5 mg L⁻¹ PO₄-P | 2% of reading |                        | Jordan et al., 2007; Wade et al., 2009 |</p>
<table>
<thead>
<tr>
<th>Electrochemistry – Ion Selective Electrodes</th>
<th>Direct potentiometry between a sensing electrode and a reference electrode</th>
<th>Nitrate</th>
<th>0-422 mg L⁻¹ NO₃-N</th>
<th>Up to 10 % or 2 mg L⁻¹, whichever is greatest, LOD 0.25 mg L⁻¹</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Ammonium</td>
<td></td>
<td>0-9000 mg L⁻¹ NH₄-N</td>
<td>Up to 10 % or 2 mg L⁻¹, whichever is greatest</td>
</tr>
<tr>
<td></td>
<td>pH</td>
<td></td>
<td>0-14 units</td>
<td>±0.2 units</td>
</tr>
</tbody>
</table>

- Calibration at least once a day due to drift
- Regular cleaning due to fouling
- May need reconditioning

Adsett and Zoerb, 1991; Le Goff et al., 2002

Merks, 1975; Toda et al., 2011

Adamchuck et al., 1999; Collings et al., 2003

* Measurement accuracy compiled from commercially available sensors. Note that accuracy under field conditions may be less than under laboratory test conditions.
<table>
<thead>
<tr>
<th>Paper</th>
<th>Study duration</th>
<th>Nutrients</th>
<th>Analysis (superscripts indicate method detailed at the foot of the table)</th>
<th>Processing time</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nutrient analysers</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bieroza &amp; Heathwaite (2015)</td>
<td>24 months</td>
<td>TP, TRP</td>
<td>In line wet chemistry analyser (2 x micromac C analysers) for TP(^4) &amp; TRP(^1)</td>
<td>TP: 50-min TRP: 10-min</td>
<td>Automated processing &amp; telemetry; 14-day lab maintenance schedule, validated by weekly grab samples.</td>
</tr>
<tr>
<td>Brinkman &amp; Hozalski (2011)</td>
<td>11 months</td>
<td>DOM</td>
<td>In-line analysis with sample pre-processing through a 3μm filter. i. DOC: portable TOC analyser ii. Composition (SUVA): UV absorbance at λ 178 to 891 nm with path length of 1cm</td>
<td>4-min intervals;</td>
<td>Validated with bi-weekly manual sampling. Analysers operational 70% (DOC) &amp; 60% (SUVA) of the time (due to clogged lines and power problems).</td>
</tr>
<tr>
<td>Carstea et al. (2009)</td>
<td>14-days</td>
<td>DOM</td>
<td>River waters sampled continuously, pre-filtered (coarse; 2mm; 0.5mm pore size) &amp; pumped (30ml/min) through a bench-top spectro-fluorometer to determine Excitation – Emission Matrices (Ex: 225-400nm; Em: 280-500nm)</td>
<td>3-min intervals</td>
<td>Hourly grab samples. Results suggest monitoring frequency of &lt;30-min required to assess DOM variability.</td>
</tr>
<tr>
<td>Cassidy &amp; Jordon (2011);</td>
<td>24-months</td>
<td>TP</td>
<td>Colorimetry: 100ml sample homogenised and 10ml delivered to analyser</td>
<td>20-min intervals</td>
<td>Daily autocalibration; underprediction of 7% of TP attributed to selective loss of heavier particles; weekly maintenance.</td>
</tr>
<tr>
<td>Duan et al. (2014)</td>
<td>36-months</td>
<td>NO(_3^-) + NO(_2^-)</td>
<td>Wet chemistry nitrate analyser; samples pre-filtered, [Envirotech NAS 3 nitrate analyser]</td>
<td>2-3 hrs</td>
<td>Automated analysis of calibration standard: every 5(^{th}) sample. Logged data accessed via internet.</td>
</tr>
<tr>
<td>Gilbert et al. (2013)</td>
<td>14 months</td>
<td>NO(_3^-), NO(_2^-), PO(_4^3-)</td>
<td>In line wet chemistry analyser; pre-processing through a 10μm filter; i. Autonomous profiling nutrient analyser (APNA) NO(_3^+)NO(_2^+), NO(_2^-), PO(_4^3-); ii. ‘Cycle PO(_4^3-)’; iii. ‘Satlantic submersible UV nutrient analyser’ NO(_3^-).</td>
<td>60 – 90 min intervals;</td>
<td>Automated processing &amp; telemetry; Validation with manual sampling and lab analyses within 24-hours.</td>
</tr>
<tr>
<td>Halliday et al. (2015)</td>
<td>(methodology from Wade et al., 2012)</td>
<td>23-months</td>
<td>TP, TRP</td>
<td>Hach Lange Phosphax Sigma: colorimetric and digestion determination of TRP(^4) and TP(^4)</td>
<td>1-hr</td>
</tr>
<tr>
<td>Halliday et al (2014)</td>
<td>24-months</td>
<td>TP, TRP, NO(_3^-), NH(_4^+), NO(_2^-)</td>
<td>Micromac C – determination of TRP(^2); NO(_2^+); NH(_4^+)</td>
<td>Intermittent pumping</td>
<td>Automated daily check with standard; reagents changed at 14-day intervals. Analysis produced 100l of waste water over 14-days requiring disposal.</td>
</tr>
<tr>
<td>Ockenden et al. (2016)</td>
<td>2 x 12-months</td>
<td>TP, TRP, NO(_3^-)</td>
<td>Hach Lange combined Sigmatax- Phosphax analyser for TP, TRP, Nitratax Plus SC probe for NO(_3^-)</td>
<td>10-min for analysis; data resolution: 30-min</td>
<td>Nitratax Plus sensor calibrated every 3-</td>
</tr>
<tr>
<td>Study</td>
<td>Timeframe</td>
<td>Methodology Details</td>
<td>Notes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>-------------------------------</td>
<td>-----------</td>
<td>----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
<td>--------------------------------------------------------------------------------------------</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jeong et al. (2012)</td>
<td>15-months</td>
<td>UV-Vis spectrophotometer [carbo::lyser s::can; Messtechnik GmbH]. Two beams for TOC &amp; DOC; 5mm path length λ 200 to 732 nm</td>
<td>Automated cleaning of sensor head every 10-mins; Site specific calibration developed from biweekly samples.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jones et al. (2014)</td>
<td>6-months</td>
<td>UV-Vis spectrolyser λ 200 to 732 nm at four sites</td>
<td>Calibration developed from 91 water samples. Weekly cleaning with 10% HCl; problems at two sites with Mn precipitation (requiring weekly cleaning)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mast et al. (2015)</td>
<td>4 years</td>
<td>Turner Cyclops-7</td>
<td>External brush wiper; data corrected for temperature and turbidity. Data validated by regular manual sampling and lab analysis.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pellerin et al (2011)</td>
<td>3-months</td>
<td>NO₃- &amp; DOM WETstar FDOM fluorometer λex/λem: 370/460nm Passing subsequently through a 0.2μm filter to a UV spectrophotometer for absorption from 217 to 240nm</td>
<td>Compensated for lamp degradation over fieldwork campaign.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Saraceno et al. (2009)</td>
<td>4-weeks</td>
<td>2 x WETstar FDOM fluorometer λex/λem: 370/460nm; one flowpath filtered, one unfiltered. FDOM: collected at 1Hz, sampled over 30-s hourly</td>
<td>Filtered system using 10μm and 0.2μm filters. Flow rates: 2L/min. System cleaned &amp; calibrated twice during 4-week field campaign.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Strohmeier et al (2013)</td>
<td>12-months</td>
<td>DOC UV-Vis; spectrophotometer (Spectro::lyser s::can; Messtechnik GmbH) UV-VIS λ: 200 to 732 nm</td>
<td>Site specific calibration developed; data verified with samples in the lab; OM quality assessed by fluorescence spectroscopy (EEM) on 116 samples</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Watras et al (2015)</td>
<td>18-months</td>
<td>DOM Turner Cyclops 7; Turner C3; SeaPoint fluorometer</td>
<td>C3 – automatic wiper; other sensors cleaned weekly</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wilson et al (2013)</td>
<td>12-months</td>
<td>DOM Turner Cyclops 7</td>
<td>Validated with lab determination of DOC on waters sampled manually (125); FDOM sensitive to temperature, composition &amp; concentration; sensor inspected monthly (biofouling during low flow)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Worrall et al (2015)</td>
<td>36-months</td>
<td>DOC UV absorbance probe (ABB AV400)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

1. phosphomolybdenum blue complexation;  
2. reaction with sulphanilamide and N-(1-naphthyl) ethylenediamine (NEDD) in acid yielding diazonium salt, detected at 525 nm;  
3. Berthelot reaction using salicylate to measure NH₄.
colorimetric determination: acid persulphate digestion