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R2MLwiN: A Package to Run MLwiN from within R

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Abstract

R2MLwiN is a new package designed to run the multilevel modeling software program MLwiN from within the R environment. It allows for a large range of models to be specified which take account of a multilevel structure, including continuous, binary, proportion, count, ordinal and nominal responses for data structures which are nested, cross-classified and/or exhibit multiple membership. Estimation is available via iterative generalized least squares (IGLS), which yields maximum likelihood estimates, and also via Markov chain Monte Carlo (MCMC) estimation for Bayesian inference. As well as employing MLwiN's own MCMC engine, users can request that MLwiN write BUGS model, data and initial values statements for use with **WinBUGS** or **OpenBUGS** (which **R2MLwiN** automatically calls via **rbugs**), employing IGLS starting values from MLwiN. Users can also take advantage of MLwiN's graphical user interface: for example to specify models and inspect plots via its interactive equations and graphics windows. **R2MLwiN** is supported by a large number of examples, reproducing all the analyses conducted in MLwiN's IGLS and MCMC manuals.

Keywords: **R2MLwiN**, MLwiN, R, **WinBUGS**, **OpenBUGS**, multilevel model, random effects model, mixed model, hierarchical linear model, clustered data, maximum likelihood estimation, Markov chain Monte Carlo estimation.

1. Introduction

In research fields as diverse as education, economics, medicine, psychology, and biology, it is commonplace to encounter data which are clustered: for example, exam results from many students across a smaller number of schools in a cross-sectional study, or clinical measurements taken repeatedly from the same individuals in a longitudinal study. Multilevel modeling is an efficient way to model such data; it accounts for the lack of independence in clustered

data, and adjusts the standard errors accordingly, whilst also opening avenues of enquiry to which more traditional multiple regression techniques are ill-suited, such as investigating group effects (e.g., Goldstein 2010; Pinheiro and Bates 2000; Raudenbush and Bryk 2002). Multilevel models (also known as mixed models, random effects models, hierarchical models, etc.) achieve this by treating the units at each level (in the above examples: students and schools, measurement occasion and individuals, respectively), as a random sample from a larger population with an assumed distribution, partitioning the residual variance between levels.

1.1. MLwiN software

The statistical software package MLwiN (Rasbash *et al.* 2009) is designed to allow researchers to specify, estimate and interpret multilevel models. It was first released, as a Windows-based program, in 1998, and is still actively-developed by the Centre for Multilevel Modelling (now at the University of Bristol). With an estimated 18,000 users worldwide, the current version (v2.36, at time of writing) was released in April 2016, and native versions of the MLwiN engine for Mac OS X and Linux have been recently made available. MLwiN allows for a variety of response types to be modeled, including continuous, binary, proportion, count, ordinal, nominal, and multivariate combinations (i.e., simultaneous equations); estimation is available via iterative generalized least squares (IGLS, Goldstein 1986), which yields maximum likelihood estimates, and also via Markov chain Monte Carlo (MCMC) estimation for Bayesian inference (Browne 2012). It supports a range of data structures, including nested, cross-classified (i.e., crossed random effects) and/or multiple membership structures (Browne *et al.* 2001). Other features include the fitting of complex level 1 variance (heteroskedastic) models, multilevel factor analysis (MCMC only; with multiple – correlated or uncorrelated – factors at each level), adjustments for measurement errors in predictors, spatial conditional autoregressive (CAR) models, autoregressive residual structures at level 1, and a selection of MCMC algorithms to increase efficiency.

Although MLwiN can be run via a macro language and the command line, this can prove unwieldy, reflecting its origins building on the earlier statistical package NANOSStat (Healy 1989). Indeed, it is likely that many users operate MLwiN via its graphical user interface (GUI); this has a number of innovative features, such as an interactive equations window, in which users can point-and-click on elements of the fully-formatted mathematical formulae representing their model to change its specification, and interactive graphical displays, allowing users to ascertain the identity of plotted points by simply clicking on them.

Funding from the UK’s Economic and Social Research Council (ESRC) has allowed MLwiN to be offered free to UK-based academics, and it is otherwise available for purchase, and as an unrestricted 30-day trial version. The Centre for Multilevel Modelling’s website (<http://www.bristol.ac.uk/cmm/>) provides details on how to obtain the software, as well as manuals and other resources offering guidance to MLwiN, and indeed to multilevel modeling in general.

1.2. R

R (R Core Team 2016) is a freely-available, open source “language and environment for statistical computing and graphics” (<https://www.R-project.org/>). It is designed to be highly extensible, as evidenced by the large number of user-authored add-on packages available on the Comprehensive R Archive Network (CRAN; <https://CRAN.R-project.org/>). In con-

trast to MLwiN, R is chiefly designed to be operated via a command line interface (CLI), although alternative user interfaces are available. Considerable support and advice is available to R users via <https://www.R-project.org/>, together with a vibrant online community of other forums and resources, as well as a range of books and manuals.

1.3. The R2MLwiN package

R2MLwiN has been designed to run MLwiN from within the R environment, combining the multilevel modeling functionality of the former with the benefits of working within the environment of the latter. For example, one can parsimoniously specify, in R, the estimation of a series of models in MLwiN, and take advantage of R's scripting language, and powerful and flexible graphing functionality, to efficiently post-process the results returned. In addition, whilst a number of sophisticated data manipulation and handling functions are available in MLwiN, R offers further flexibility, including the import of data saved in a wide range of formats. Furthermore, since the recently-released native versions of the MLwiN engine for Mac OS X and Linux only offer an MLwiN macro interface, **R2MLwiN** offers a convenient means of interacting with them. More generally, performing analyses via the use of command line scripts can facilitate faithful documentation and thus aid reproducibility.

As with all packages available on CRAN, **R2MLwiN** has supporting help files, and also has a supporting website maintained by **R2MLwiN**'s authors (<http://www.bristol.ac.uk/cmm/software/r2mlwin/>). In addition, **R2MLwiN** comes with a large number of demos: R scripts replicating all the analyses conducted in the user's guide to MLwiN (Rasbash *et al.* 2012), which describes model-fitting via iterative generalized least squares (IGLS) estimation (which yields maximum likelihood estimates), and the guide to MCMC estimation in MLwiN (Browne 2012). The following will return a list of all available demos (note: in all the example script which follows, R> denotes the R prompt, whilst +, if appearing at the start of a line, indicates it is a continuation of the line above):

```
R> library("R2MLwiN")
R> demo(package = "R2MLwiN")
```

Note that on loading **R2MLwiN**, the default path to MLwiN will be stated in the text returned. This can be changed away from the default via `options(MLwiN_path = "path/to/MLwiN vX.XX/")`. A specific demo can be run via:

```
R> demo("MCMCGuide03")
```

In the following sections we work through a variety of examples chosen firstly to illustrate the fundamentals of working with **R2MLwiN**, including for those relatively inexperienced in R, MLwiN and/or multilevel modelling in general, and secondly to highlight features which may be of particular interest to R users wishing to fit multilevel models, such as functionality not commonly-supported by other R packages. We begin with an example using IGLS, and then move on to consider MCMC. The models start fairly simple so that we can familiarize the user with the syntax used in **R2MLwiN** while in later sections we describe more complex models.

2. Fitting a 2-level continuous response model via IGLS

For our first example, we will fit a 2-level continuous response model using IGLS, employing an educational dataset available as the sample dataset `tutorial` in both MLwiN and R2MLwiN. This is a subset of data from a much larger dataset of examination results from six inner-London Education Authorities (school boards). The original analysis (Goldstein *et al.* 1993) sought to establish whether some secondary schools had better student exam performance at 16 than others, *after* taking account of variations in the characteristics of students when they started secondary school; i.e., the analysis investigated the extent to which schools ‘added value’ (with regard to exam performance), and then examined what factors might be associated with any such differences.

```
R> data("tutorial")
```

The variables we will analyze in the following examples are summarized in Table 1 (although you can view descriptions of all the variables in the original dataset by typing `?tutorial`).

We will start with an example of the simplest multilevel model one can fit: a 2-level variance components model with a continuous response. Such models partition the variance in the response variable between the levels specified in the model. We will choose `normexam` as our response variable for `student` i in `school` j ; see Equation 1.

$$\begin{aligned} \text{normexam}_{ij} &= \beta_0 + u_j + e_{ij} \\ u_j &\sim N(0, \sigma_u^2) \\ e_{ij} &\sim N(0, \sigma_e^2) \end{aligned} \tag{1}$$

Here, u_j corresponds to the `school`-level random effect whilst e_{ij} is the `student`-level residual error; u_j and e_{ij} are assumed to be independent of one another and normally-distributed with zero means and constant variances σ_u^2 and σ_e^2 . The model thus partitions the variance in `normexam` between that attributable to schools (σ_u^2), corresponding to departures of the school means from the overall mean (β_0), and that attributable to students within schools (σ_e^2), corresponding to departures of students’ `normexam` scores from the mean of the school they attend.

We can fit this model as follows:

```
R> F1 <- normexam ~ 1 + (1 | school) + (1 | student)
R> (VarCompModel <- runMLwiN(Formula = F1, data = tutorial))
```

Variable	Description
<code>school</code>	Numeric school identifier.
<code>student</code>	Numeric student identifier.
<code>normexam</code>	Students’ exam score at age 16, normalized to have approximately a standard normal distribution.
<code>standlrt</code>	Student’s score at age 11 on the London reading test (LRT), standardized using Z-scores.

Table 1: Variables in the `tutorial` dataset, as modeled in the worked example.

Here we have created an object, `VarCompModel`, to which we have assigned a call (with appropriate arguments) to **R2MLwiN**'s `runMLwiN` function. In this example the `Formula` and `data` arguments are the only ones explicitly declared. In the case of the `Formula`, `normexam` has been specified as the response variable, since it is to the left of the tilde (`~`), and only an intercept is included in the fixed part of the model. Note that the intercept is not included by default (this is keeping with the manner in which models are specified in `MLwiN`), and so is explicitly added by including `1` to the right of the tilde. The random part of the model is specified in sets of parentheses arranged in descending order with respect to their hierarchy. So here we have specified that the coefficient of the intercept is allowed to randomly-vary at level 2 (`1 | school`) and level 1 (`1 | student`). Note that the variable containing the level 1 ID needs to be explicitly specified unless the model is a discrete response model (in which case it should not be specified). See Table 2 (plus later examples) for details of how to specify the `Formula` argument when modeling other distributions, and also Section 10 for an example using categorical explanatory variables.

Other than nominating the `data.frame` containing the variables to be modeled in the `data` argument, we accept the defaults for `runMLwiN`'s other arguments (see `?runMLwiN`). These include the distribution to be modeled (defaulting to `D = "Normal"`; see Table 2 for other distributions, many of which are explored in later examples) and also an argument pertaining to estimation options; this defaults to IGLS estimation, which is denoted by `estoptions = list(EstM = 0)`. The argument `estoptions` is a list with a large number of possible terms – some are covered in later examples, but a more comprehensive account is provided in the relevant **R2MLwiN** help files. We also accept the default `workdir = tempdir()`, which indicates the output files are to be saved in the temporary directory.

Once this command has been entered, functions within the **R2MLwiN** package will then take this input and create an `MLwiN` macro file; `MLwiN` is then called and executes the macro script, and the output is returned to R for post-processing, such as producing the following output table (which can be reproduced via e.g., `summary(VarCompModel)`):

```

-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-***-
MLwiN (version: 2.36)  multilevel model (Normal)
      N min      mean max N_complete min_complete mean_complete max_complete
school 65    2 62.44615 198          65             2      62.44615          198
Estimation algorithm:  IGLS          Elapsed time : 0.12s
Number of obs: 4059 (from total 4059) The model converged after 3 iterations.
Log likelihood:          -5505.3
Deviance statistic:     11010.6

```

```

-----
The model formula:
normexam ~ 1 + (1 | school) + (1 | student)
Level 2: school      Level 1: student

```

```

-----
The fixed part estimates:
      Coef.   Std. Err.      z    Pr(>|z|)    [95% Conf.  Interval]
Intercept -0.01317    0.05363   -0.25    0.806    -0.11827    0.09194
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```

-----
The random part estimates at the school level:

```

D	Formula	Where <link> can equal ...
"Normal"	$\langle y1 \rangle \sim 1 + \langle x1 \rangle + \dots + (1 \langle L2 \rangle) + (1 \langle L1 \rangle)$	(identity link assumed)
"Poisson"	$\langle \text{link} \rangle(\langle y1 \rangle) \sim 1 + \text{offset}(\langle \text{offs} \rangle) + \langle x1 \rangle + \dots + (1 \langle L2 \rangle)$	log
"Negbinom"	$\langle \text{link} \rangle(\langle y1 \rangle) \sim 1 + \text{offset}(\langle \text{offs} \rangle) + \langle x1 \rangle + \dots + (1 \langle L2 \rangle)$	log
"Binomial"	$\langle \text{link} \rangle(\langle y1 \rangle, \langle \text{denom} \rangle) \sim 1 + \langle x1 \rangle + \dots + (1 \langle L2 \rangle)$	logit, probit, cloglog
"Unordered Multinomial"	$\langle \text{link} \rangle(\langle y1 \rangle, \langle \text{denom} \rangle, \langle \text{ref_cat} \rangle) \sim 1 + \langle x1 \rangle + \dots + (1 \langle L2 \rangle)$	logit
"Ordered Multinomial"	$\langle \text{link} \rangle(\langle y1 \rangle, \langle \text{denom} \rangle, \langle \text{ref_cat} \rangle) \sim 1 + \langle x1 \rangle + \langle x2 \rangle[\langle \text{common} \rangle] + \dots + (1[\langle \text{common} \rangle] \langle L3 \rangle) + (1 \langle L2 \rangle)$	logit, probit, cloglog
"Multivariate Normal"	$c(\langle y1 \rangle, \langle y2 \rangle, \dots) \sim 1 + \langle x1 \rangle + \langle x2 \rangle[\langle \text{common} \rangle] + \dots + (1[\langle \text{common} \rangle] \langle L3 \rangle) + (1 \langle L2 \rangle) + (1 \langle L1 \rangle)$	(identity link assumed)
c("Mixed", "Binomial", "Normal")	$c(\langle \text{link} \rangle(\langle y2 \rangle, \langle \text{denom} \rangle), \langle y1 \rangle, \dots) \sim 1 + \langle x1 \rangle + \langle x2 \rangle[\langle \text{common} \rangle] + \dots + (1[\langle \text{common} \rangle] \langle L3 \rangle) + (1 \langle L2 \rangle) + (1[\langle \text{Normal_resp} \rangle] \langle L1 \rangle)$	logit‡, probit, cloglog‡
c("Mixed", "Poisson", "Normal")‡	$c(\langle \text{link} \rangle(\langle y2 \rangle), \langle y1 \rangle, \dots) \sim 1 + \text{offset}(\langle \text{offs} \rangle) + \langle x1 \rangle + \langle x2 \rangle[\langle \text{common} \rangle] + \dots + (1[\langle \text{common} \rangle] \langle L3 \rangle) + (1 \langle L2 \rangle) + (1[\langle \text{Normal_resp} \rangle] \langle L1 \rangle)$	log

Table 2: A summary of options for the `Formula` argument in **R2MLwiN**. They assume an intercept is added (which needs to be explicitly specified, here via the addition of 1). `<link>` denotes the link function, `<y1>`, `<y2>`, etc. represent response variables, `<denom>` denotes the denominator (optional; if not specified, a constant of ones is used as denominator), `<offs>` the offset (optional), `<L2>`, `<L1>`, etc. the variables containing the level 2 and level 1 identifying codes, and `<ref_cat>` represents the reference category of a categorical response variable (optional: if unspecified the lowest level of the factor is used as the reference category). Explanatory variables are specified as e.g., `<x1> + <x2>`. For "Ordered Multinomial", "Multivariate Normal" and "Mixed" responses, `[<common>]` indicates a common coefficient (i.e., the same for each category) is to be fitted; here `<common>` takes the form of a numeric identifier indicating the responses for which a common coefficient is to be added (e.g., `[1:5]` to fit a common coefficient for categories 1 to 5 of a 6-point ordered variable, `[1]` to fit a common coefficient for the response variable specified first in the `Formula` object for a "Mixed" response model, etc.). Otherwise a separate coefficient (i.e., one for each category) is added. `<Normal_resp>` indicates the position (as an integer) the Normal response(s) appears in the formula (2 in the examples in the table). For "Mixed" response models, the `Formula` arguments need to be grouped in the order the distributions are listed in D. Note that, when using IGLS, quasi-likelihood methods are used for discrete response models (and not, for example, adaptive quadrature). ‡ denotes IGLS only.

	Coef.	Std. Err.
var_Intercept	0.16863	0.03245

The random part estimates at the student level:

	Coef.	Std. Err.
var_Intercept	0.84776	0.01897

The first few lines returned, not reproduced here (and omitted from the output below too), provide information on how estimation is progressing during the model fit (such as the iteration number), as well as a few miscellaneous MLwiN commands. Once convergence has been achieved then the summary is returned as above, confirming first the version of MLwiN used, the distribution chosen, and then some summary statistics concerning the level 2 units: namely that there were 65 schools containing a mean of 62.4 students, ranging from a minimum of 2 to a maximum of 198 students (separate summary statistics, with the suffix `_complete`, are displayed alongside pertaining to complete cases only; unless a level 2 unit has data missing for every level 1 unit, N will equal `N_complete`). Otherwise, the estimation method is confirmed, the time elapsed between R exporting the data/macros and the results being returned, the number of (complete) observations from the total in the dataset (this dataset has 4059 students, with no missing data for any of the modeled variables), confirmation that the model converged after 3 iterations, the log likelihood, and finally the deviance statistic ($-2 \times \log \text{likelihood}$) which can be used to compare the fit of ‘nested’ models via likelihood ratio tests (see below).

Beneath that, there is a reiteration of the model formula/levels we specified earlier, followed by the model estimates. First we see the coefficient estimates for the fixed part of the model, which in this case consists only of the intercept, indicating that the overall mean of `normexam` is -0.013 (which is close to zero since it has been standardized), with a standard error of 0.054, together with the corresponding z score, its p value, and confidence intervals (although these latter statistics are of little substantive interest in this particular instance).

Next we have the estimates (with standard errors) for the random part of the model, indicating that the `school` means are distributed around the overall mean with an estimated variance of 0.169 ($\hat{\sigma}_u^2$), with the (within-school) `student` scores distributed around their `school` mean with an estimated variance of 0.848 ($\hat{\sigma}_e^2$).

2.1. Conducting a likelihood ratio test

If we wish to investigate whether there are significant school differences in `normexam` (i.e., whether it is necessary to take account of clustering at the `school`-level or not), we can conduct a likelihood ratio test: subtracting the deviance statistic from the current 2-level model from the corresponding value from the simpler single-level model (Equation 2) and comparing this against a χ^2 distribution with the appropriate degrees of freedom (in this case 1, since the addition of only one parameter distinguishes the two models).

$$\begin{aligned} \text{normexam}_i &= \beta_0 + e_i \\ e_i &\sim N(0, \sigma_e^2) \end{aligned} \tag{2}$$

We do so here using the generic S4 method `logLik` included in the `mlwinfitIGLS-class`

(IGLS model fit) object; this allows us to conduct a likelihood ratio test using the `lrtest` function, which is part of the `lmtest` package (Zeileis and Hothorn 2002):

```
R> F2 <- normexam ~ 1 + (1 | student)
R> OneLevelModel <- runMLwiN(Formula = F2, data = tutorial)
R> if (!require(lmtest)) install.packages("lmtest")
R> lrtest(OneLevelModel, VarCompModel)
```

Likelihood ratio test

```
Model 1: OneLevelModel
Model 2: VarCompModel
  #Df  LogLik Df  Chisq Pr(>Chisq)
1    2 -5754.7
2    3 -5505.3  1 498.72 < 2.2e-16 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

As such, we can see there are highly-significant effects of `school` on pupils' `normexam` scores.

2.2. Calculating the variance partition coefficient (VPC)

If we wanted to find out how much of the variance in `normexam` is due to differences between schools, and how much is due to differences between students within schools, then we could calculate the variance partition coefficient using the relevant slots for this `S4` class object to pull out the estimates we need for the calculation; in this instance we use the slot `"RP"`, which stands for “random part” (typing `slotNames(VarCompModel)` will list all available slots):

```
R> print(VPC <- VarCompModel["RP"][["RP2_var_Intercept"]] /
+ (VarCompModel["RP"][["RP1_var_Intercept"]] +
+ VarCompModel["RP"][["RP2_var_Intercept"]]))

[1] 0.1659064
```

i.e., approximately 17% of the total variance in `normexam` is attributable to differences between schools. (Note that calculating the VPC is less straightforward for random coefficients and discrete response models; see, e.g., Goldstein *et al.* 2002.)

3. Storing residuals

By default, **R2MLwiN** does not store residuals (at any level), but it is straightforward to request it to do so by simply toggling the value of `resi.store`, in the list of `estoptions`, as follows (see Section 8, as well as `?runMLwiN`, for more advanced options regarding saving residuals):

```
R> VarCompResid <- runMLwiN(Formula = F1, data = tutorial,
+   estoptions = list(resi.store = TRUE))
```

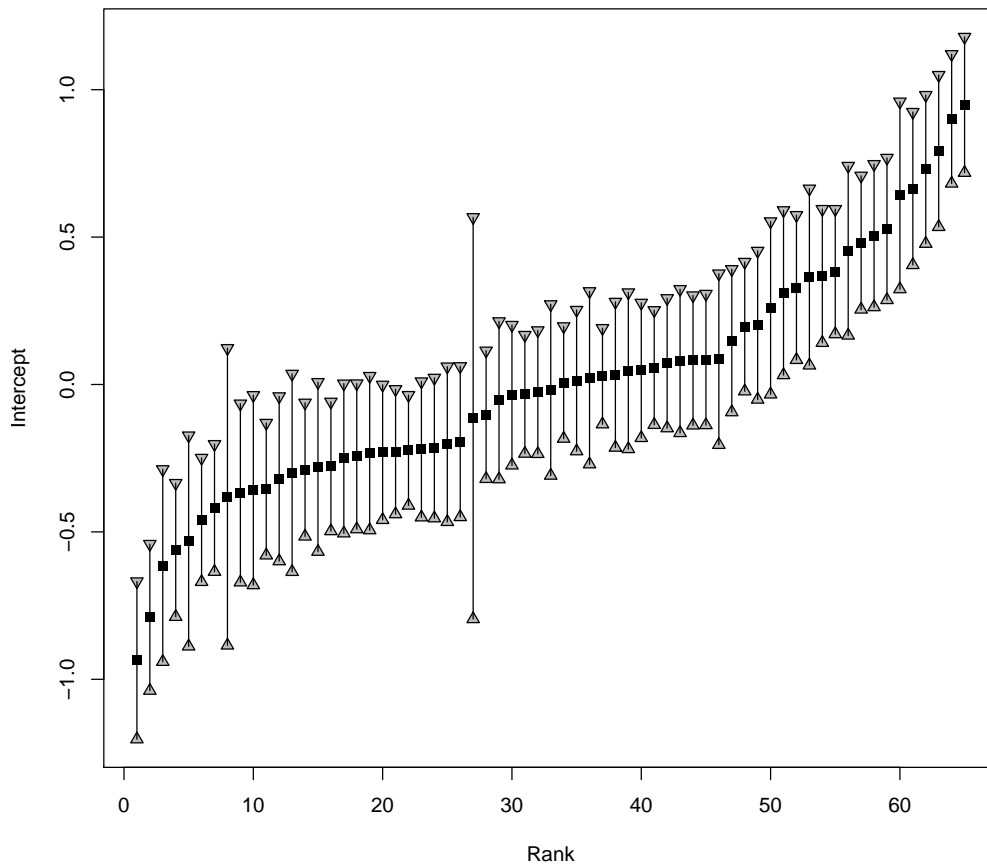


Figure 1: Caterpillar plot of level 2 residuals produced by **R2MLwiN**'s caterpillar function.

We can then, for instance, plot the residuals using **R2MLwiN**'s `caterpillar` function, a wrapper for the `plot` function with the addition of error bars, yielding the plot displayed in Figure 1; see the **R2MLwiN** demo `MCMCGuide04` for an alternative choice of caterpillar plot:

```
R> residuals <- VarCompResid@residual$lev_2_resi_est_Intercept
R> residualsCI <- 1.96 * sqrt(VarCompResid@residual$lev_2_resi_var_Intercept)
R> residualsRank <- rank(residuals)
R> rankno <- order(residualsRank)
R> caterpillar(y = residuals[rankno], x = 1:65,
+   qtlow = (residuals - residualsCI)[rankno],
+   qtup = (residuals + residualsCI)[rankno],
+   xlab = 'Rank', ylab = 'Intercept')
```

4. A note on `debugmode` and `show.file`

If we had included `debugmode = TRUE` within the list of `estoptions` when fitting our variance components model, above, this would have left the **MLwiN** application open if running via Windows or in Wine (Wine Development Team 2015) on other platforms, with our variables uploaded, the equation window displayed, and the starting values (in blue) of our specified

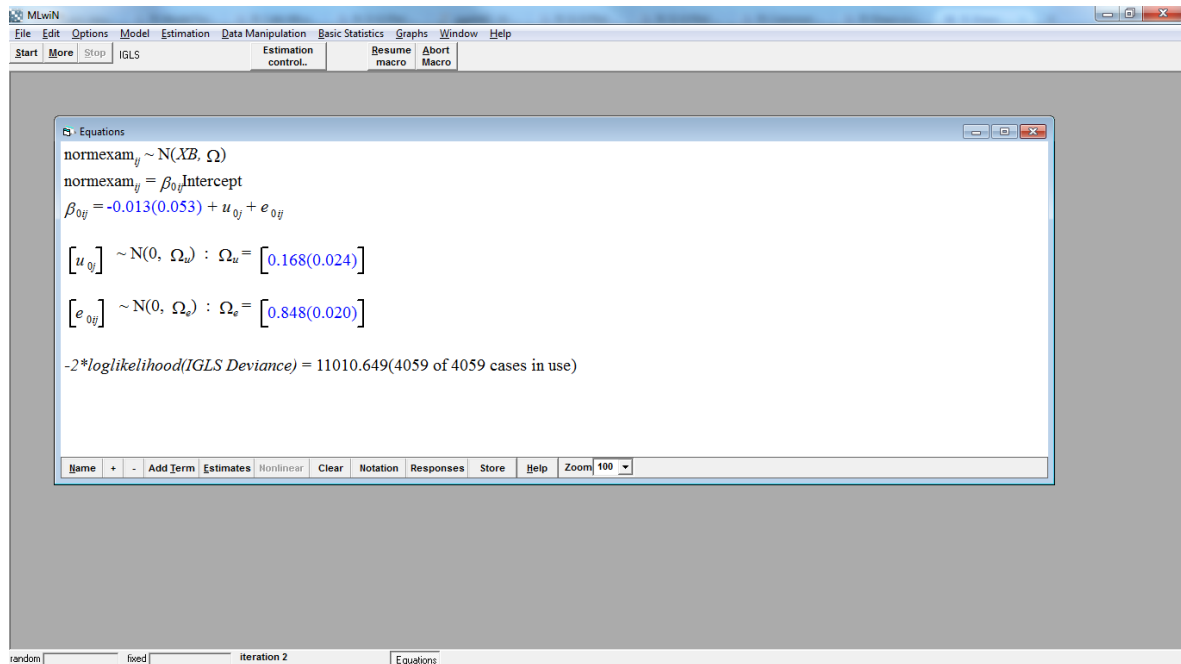


Figure 2: The MLwiN application is left open, with the interactive Equations window displayed, if `debugmode` is specified as `TRUE`.

model; see Figure 2. Pressing the ‘Resume macro’ button, within MLwiN, results in the model iterating to convergence, at which point the estimates turn green. Pressing the button again returns a dialogue box inviting the user to either close the application, or continue working in the MLwiN GUI; in either case estimates from the model as fitted up to that point are returned to R once the MLwiN application is closed (closing the application via another method may result in model estimates not being returned to R, and an error being displayed as a result). Working with MLwiN’s GUI allows us to employ some of its useful features, such as the interactive equations and graph windows, as well as providing a useful aid to trouble-shooting in the case of model misspecification (for further guidance see [Rasbash *et al.* 2012](#); [Browne 2012](#)).

Including `show.file = TRUE` within our list of `estoptions` would display the MLwiN macro file produced by **R2MLwiN** (`clean.files = FALSE` may also need to be added to the list of `estoptions`, depending on the R interface you are using); an example taken from our variance components model, above, can be found in the supplementary materials; for further information about MLwiN commands see [Rasbash *et al.* \(2003\)](#). This macro file is also saved in the working directory, as specified in the `runMLwiN` argument `workdir`, together with a range of other outputs, such as a data file in *Stata* format (i.e., `*.dta`; these can be imported by both MLwiN and R); these are deleted from the working directory once MLwiN closes, so running in `debugmode` provides an opportunity to access them.

5. Fitting a 2-level continuous response model via MCMC

We fitted our variance components model, above, using the default estimation method of

IGLS, but we can fit the same model using MCMC methods by simply toggling the value of `EstM` (away from the default) as demonstrated below. `MLwiN`'s MCMC engine uses Gibbs sampling for steps in the algorithm where the conditional posterior distribution has a standard form (as is the case for all parameters in the current model), and, if not, uses single site random walk Metropolis sampling with a normal proposal distribution and adaptive procedure (Browne and Draper 2000).

R has, of course, a variety of other packages which fit models using MCMC methods (e.g., Park *et al.* 2016), for example the `MCMCglimm` package written by Jarrod Hadfield (Hadfield 2010) covers many of the same models as `MLwiN` and also has an impressively efficient MCMC implementation, particularly for normal response models. However, for non-normal models it does require that they contain over-dispersion terms for each observation, in part so that it can efficiently estimate the model. This is less of an issue for count data, but for binary responses can be problematic as it is not clear in such models that over-dispersion at the observation level makes sense (Skrondal and Rabe-Hesketh 2007). Gelman and Hill (2007) cover many aspects of multilevel modelling within R, including, from an MCMC perspective, how to write an MCMC algorithm directly in the R language, and to call `WinBUGS` from R via `R2WinBUGS` (Sturtz *et al.* 2005), which requires the user to write their own model code to send to `WinBUGS`, whilst `R2MLwiN` generates these files, using (by default) IGLS starting values, on behalf of the user; see Section 12.

Here we fit the model using the default MCMC estimation settings, but we would ultimately need to investigate whether these were appropriate or not:

```
R> (VarCompMCMC <- runMLwiN(Formula = F1, data = tutorial,
+   estoptions = list(EstM = 1)))
```

```

-----
MLwiN (version: 2.36) multilevel model (Normal)
      N min      mean max N_complete min_complete mean_complete max_complete
school 65   2 62.44615 198           65             2       62.44615       198
Estimation algorithm: MCMC      Elapsed time : 1.15s
Number of obs:4059 (from total 4059) Number of iter:5000 Chains:1 Burn-in:500
Bayesian Deviance Information Criterion (DIC)
Dbar      D(thetabar)      pD      DIC
10850.046  10790.013  60.034  10910.080
-----

The model formula:
normexam ~ 1 + (1 | school) + (1 | student)
Level 2: school      Level 1: student
-----

The fixed part estimates:
      Coef. Std. Err.      z  Pr(>|z|)  [95% Cred. Interval]  ESS
Intercept -0.01212  0.05410  -0.22  0.8227  -0.11977  0.08958  189
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
-----

The random part estimates at the school level:
      Coef. Std. Err.  [95% Cred. Interval]  ESS
var_Intercept  0.17629  0.03606  0.11876  0.26114  3212

```

 The random part estimates at the student level:

	Coef.	Std. Err.	[95% Cred.	Interval]	ESS
var_Intercept	0.84862	0.01887	0.81254	0.88678	5016

Here we see, towards the top of the output, that the chain has run for 5000 main iterations; this is the default in MLwiN, along with a burn-in of 500, a thinning factor of 1, and a random number seed of 1. We have also run just one chain (the default); indeed, whilst it is not possible to run more than one chain via the MLwiN GUI, it is via **R2MLwiN**, which can run multiple chains in parallel. If we wished not to use MLwiN's defaults, then such parameters can be specified in the `mcmcMeth` list within the `estoptions` argument, e.g., `estoptions = list(EstM = 1, mcmcMeth = list(burnin = 1000, nchains = 3, iterations = 10000, thinning = 10, seed = 1:3))`.

Below that we have the deviance information criterion (DIC) model-fit statistic ([Spiegelhalter et al. 2002](#)), and the quantities from which it is derived:

- \bar{D} (`Dbar`): Average deviance across all (post-burnin) iterations.
- $D(\bar{\theta})$ (`D(thetabar)`): Deviance evaluated at the estimated means of the unknown parameters θ .
- pD : The effective number of parameters, computed as $\bar{D} - D(\bar{\theta})$.
- DIC : Computed as $D(\bar{\theta}) + 2pD$: i.e., it is a measure of goodness of model fit which adjusts for model complexity, thus allowing models to be compared: models with smaller DIC values are preferred to those with large values, with differences of 5 or more considered substantial (e.g., [Lunn et al. 2012](#)).

Finally, following the model formula, we have the model estimates, which are largely similar to those from IGLS, and include 95% credible intervals (i.e., the 2.5th and 97.5th quantiles of each chain) for both the fixed and random part of the model. Note that, for the fixed part of the model, users can toggle between displaying the z score and its associated (two-tailed) p value (i.e., assuming normality), and a one-sided Bayesian p value (see Section 6 for an example).

The accompanying statistics include the ESS, or effective sample size ([Kass et al. 1998](#)), i.e., the number of iterations necessary to obtain these estimates if the sample were independent and identically-distributed. As such, a low ESS (compared to the number of iterations the chain has actually run for) would indicate a strongly positively autocorrelated chain, suggesting it may need to be run longer and/or sampled in a different manner, or other alternative MCMC methodologies could be employed to aid efficiency (see Section 11).

As well as the ESS, other MCMC diagnostic aids available in the **R2MLwiN** package include trajectory plots, as well as a larger suite of diagnostics available via the `sixway` function. We will demonstrate the latter when plotting the chain for the variance partition coefficient (VPC) in the following section; with regard to the former, however, entering

```
R> trajectories(VarCompMCMC)
```

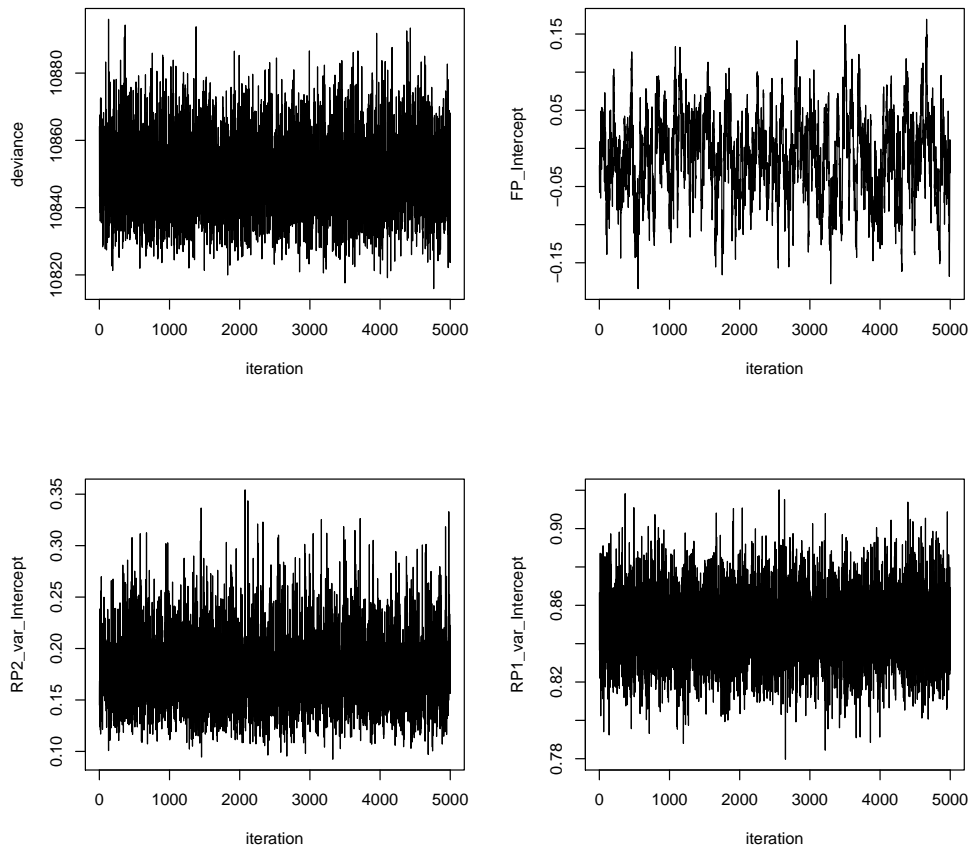


Figure 3: Plots of MCMC chain trajectories produced by **R2MLwiN**'s `trajectories` function.

yields the plots presented in Figure 3, namely the trajectories for the deviance, the intercept in the fixed part of the model (`FP_Intercept`), and the variances at level 2 (`RP2_var_Intercept`), and level 1 (`RP1_var_Intercept`) associated with the random part of the model.

5.1. Calculating the variance partition coefficient (VPC)

For our IGLS model fitted earlier, we calculated the VPC based on a single set of point estimates; in the case of MCMC, however, we have chains for σ_u^2 and σ_e^2 , and so can derive a chain for the VPC, as follows:

```
R> VPC_MCMC <- VarCompMCMC["chains"][,"RP2_var_Intercept"] /
+   (VarCompMCMC["chains"][,"RP1_var_Intercept"] +
+   VarCompMCMC["chains"][,"RP2_var_Intercept"])
R> sixway(VPC_MCMC, name = "VPC")
```

As you can see (Figure 4), using **R2MLwiN**'s `sixway` function we obtain a range of useful statistics we can use to describe this function, including mean, mode, and interval estimates (e.g., the 95% Bayesian credible interval runs from 0.123 to 0.235), as well as a range of other

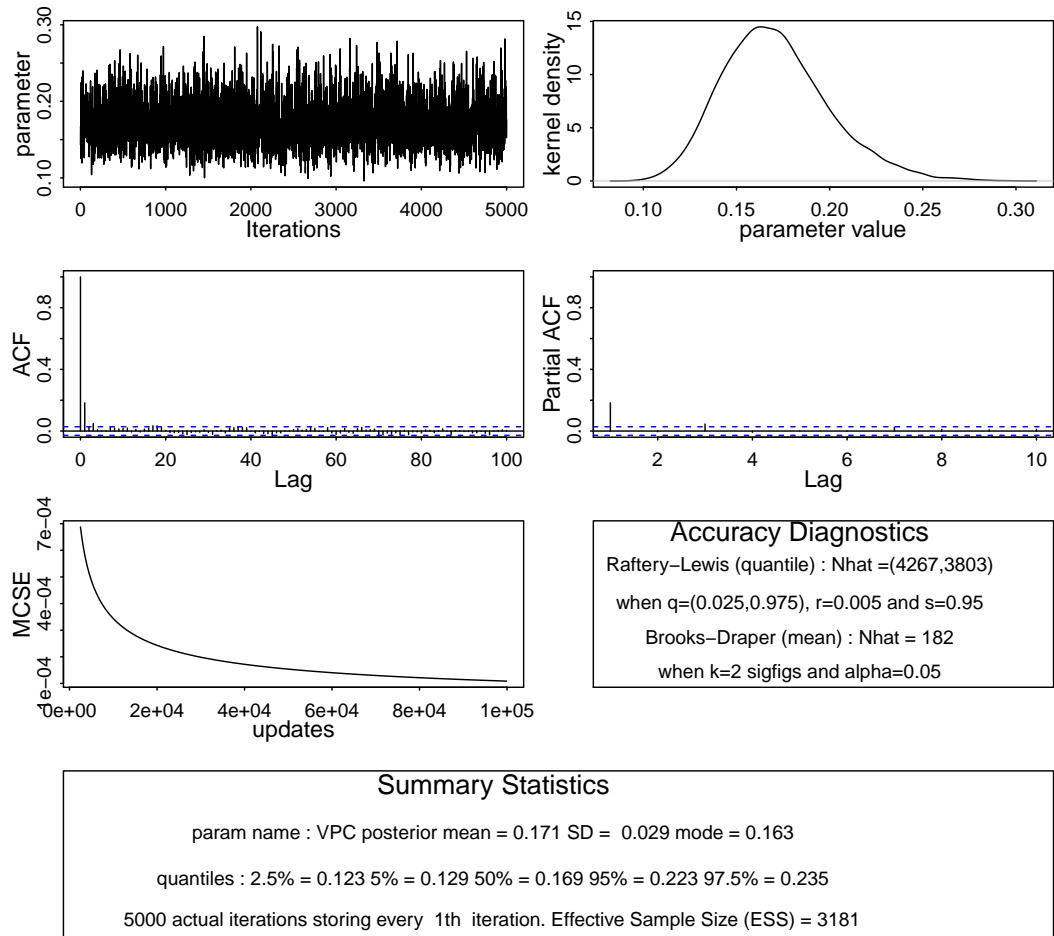


Figure 4: An example of diagnostic plots for MCMC chains produced by **R2MLwiN**'s `sixway` function.

charts and diagnostics: in addition to the trajectory plot (top-left), we also have a kernel density plot of the posterior distribution (top-right), the Raftery-Lewis diagnostic (Raftery and Lewis 1992) – indicating how long the chain should run to estimate the 0.025 and 0.975 quantiles (respectively) to a given accuracy (see defaults for the `raftery.diag` function in the `coda` package), and also the Brooks-Draper diagnostic – indicating the number of iterations the chain needs to run to estimate the mean to two significant places; Chapter 3 of Browne (2012) describes these, and the other diagnostics returned, in more detail.

The chains returned by **R2MLwiN**, as `mcmc` objects, are also, of course, available for analysis by a range of other packages and functions in R, including the many useful functions available in the `coda` package (Plummer *et al.* 2006).

6. Adding a predictor to the fixed part of a model

If we wish to see if a predictor, e.g., students' exam performance at age 11, appreciably

improves the fit of the model, we can add it to the fixed part, as in Equation 3.

$$\begin{aligned} \text{normexam}_{ij} &= \beta_0 + \beta_1 \text{standlrt}_{ij} + u_j + e_{ij} \\ u_j &\sim N(0, \sigma_u^2) \\ e_{ij} &\sim N(0, \sigma_e^2) \end{aligned} \quad (3)$$

```
R> F3 <- normexam ~ 1 + standlrt + (1 | school) + (1 | student)
R> (standlrtMCMC <- runMLwiN(Formula = F3, data = tutorial,
+   estoptions = list(EstM = 1)))
```

In the resulting output (truncated below) we see its coefficient estimate is positive and much greater than its standard error, indicating that it is a highly significant predictor, as reflected in its 95% credible interval, which excludes zero:

The fixed part estimates:

	Coef.	Std. Err.	z	Pr(> z)	[95% Cred. Interval]	ESS
Intercept	0.00514	0.04246	0.12	0.9036	-0.07933 0.08896	211
standlrt	0.56321	0.01250	45.05	0 ***	0.53862 0.58791	4060

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Note that if we wish to instead view one-sided Bayesian p values – i.e., the proportion of chain iterations which are of the opposite sign to the chain mean (the coefficient estimate in our output) – then we can request this as follows; here the output (again, truncated) indicates that the proportion of chain iterations which were below zero for the `Intercept` was 0.444, whilst none of the 5000 iterations were below zero for `standlrt`:

```
R> print(standlrtMCMC, z.ratio = FALSE)
```

The fixed part estimates:

	Coef.	Std. Err.	pMCMC(1-sided)	[95% Cred. Interval]	ESS
Intercept	0.00514	0.04246	0.444	-0.07933 0.08896	211
standlrt	0.56321	0.01250	0	0.53862 0.58791	4060

We also see a large reduction in the DIC in this model):

```
R> VarCompMCMC["BDIC"][["DIC"]] - standlrtMCMC["BDIC"][["DIC"]]
```

```
[1] 1640.952
```

7. Priors, starting values, and random seeds

Browne (2012) describes in detail the priors, starting values, and random seeds used by default by MLwiN's MCMC engine, and how to modify them to suit users' own needs, so we will not fully reiterate those details here, other than providing the following few examples by means of illustration of how to do so via **R2MLwiN**.

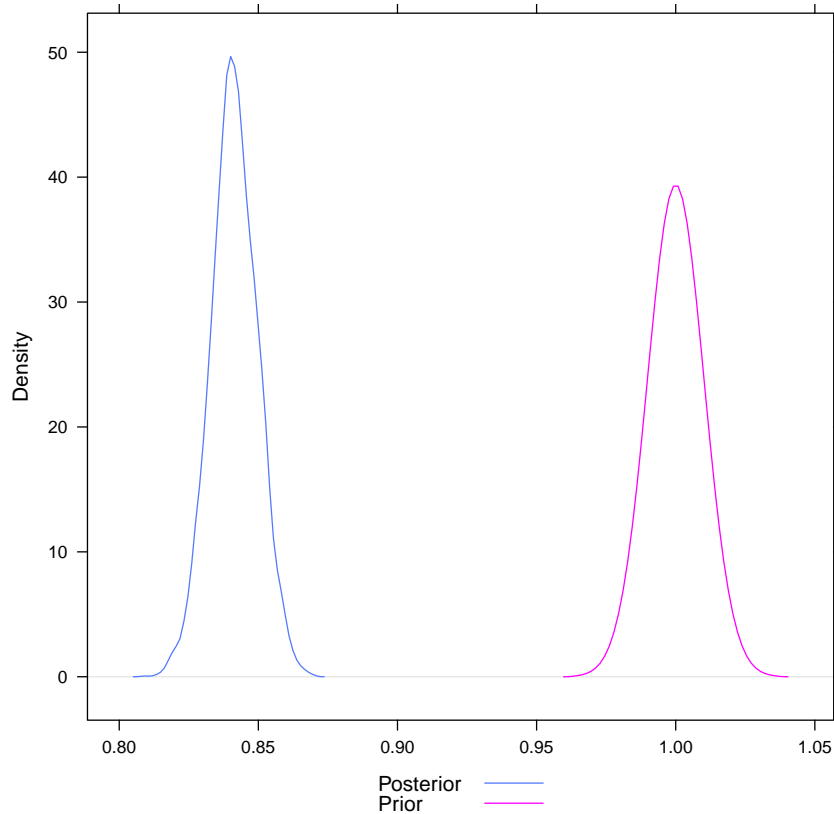


Figure 5: A `densityplot` of the prior and posterior distributions for `standlrt`.

For fixed parameters MLwiN employs, by default, an improper uniform prior $p(\beta) \propto 1$, but conjugate informative priors can readily be specified as a normal distribution with a mean and standard deviation as determined by the user. Let us assume we have gleaned, from prior work, some knowledge leading us to propose a prior for `standlrt` with a mean of 1 and standard deviation of 0.01; we can pass this information onto `priorParam`, listed within `mcmcMeth`, itself listed within `estoptions`, by specifying (via `fixe`) that we wish to construct a proper normal prior for the term in the fixed part of the model called `standlrt`, with `c(mean, sd)`. See Figure 5 for a plot produced using the `lattice` package (Sarkar 2008), automatically loaded with **R2MLwiN**, indicating a strong prior which disagrees with the data.

```
R> informpriorMCMC <- runMLwiN(Formula = F3, data = tutorial,
+   estoptions = list(EstM = 1,
+   mcmcMeth = list(priorParam = list(fixe = list(standlrt = c(1, 0.01))))))
R> PlotDensities <- data.frame(
+   PostAndPrior = c(informpriorMCMC["chains"][, "FP_standlrt"],
+   qnorm(seq(1 / 5000, 1, by = 1 / 5000), mean = 1, sd = 0.01)),
+   id = c(rep("Posterior", 5000), rep("Prior", 5000)))
R> densityplot(~ PostAndPrior, data = PlotDensities, groups = id, ref = TRUE,
+   plot.points = FALSE, auto.key = list(space = "bottom"), xlab = NULL)
```

Here, by means of illustration, we have described changing the prior for a parameter in the fixed part of the model, but **R2MLwiN** allows conjugate priors for other parameters to

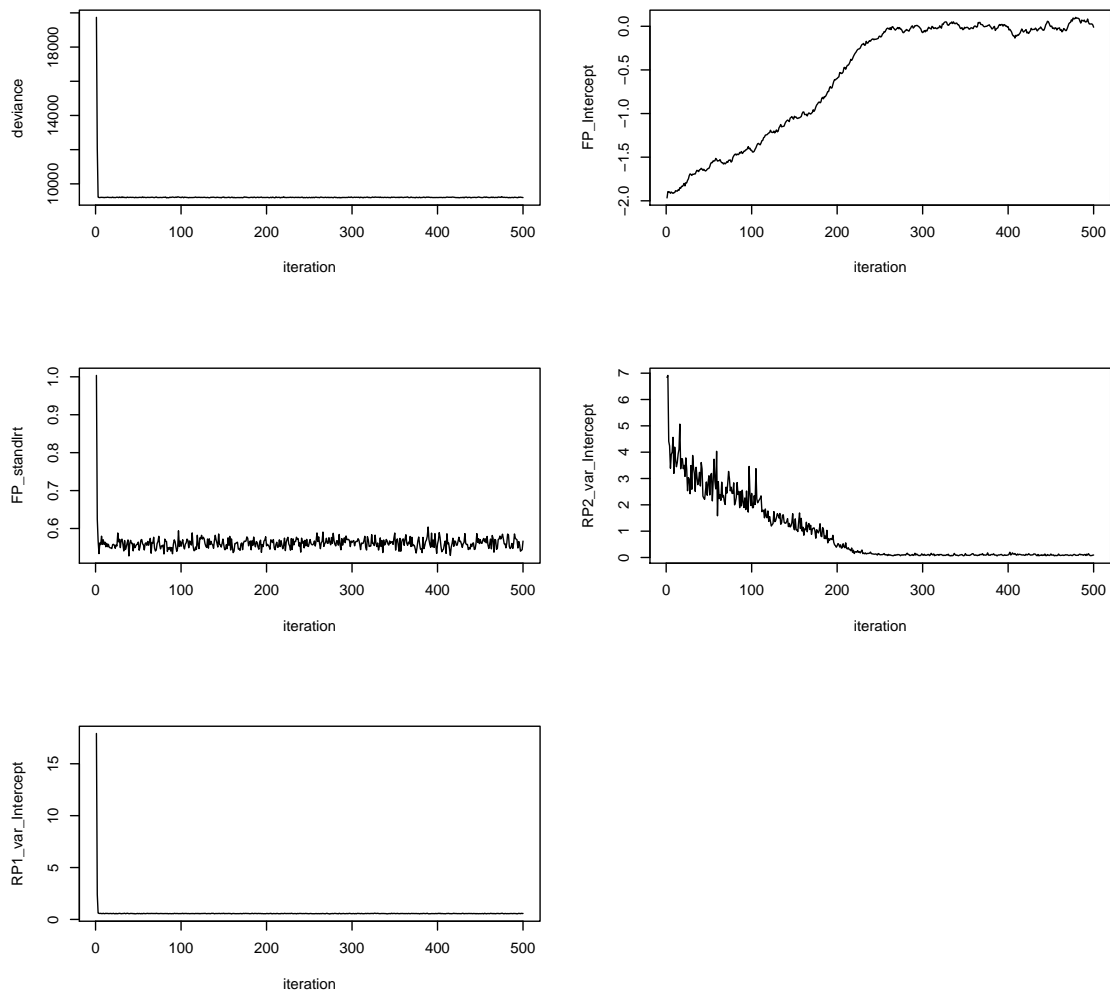


Figure 6: Chain trajectories from a model fit with custom (non-IGLS) starting values.

be modified as well, including Wishart or gamma priors for the covariance matrix, or scalar variance, in the random part of the model; see `?runMLwiN` or `?prior2macro` for further details. By default, MLwiN's MCMC engine uses IGLS estimates as starting values for each parameter; this is one of the reasons its MCMC engine converges comparatively quickly which is an advantage over for example **WinBUGS**. However, if we wished to change the starting values, we can easily do so via the `estoptions` argument: specifically the `startval` list. Here we have specified starting values of -2 and 5 for the coefficient estimates of the `Intercept` and `standlrt`, respectively, in the fixed part of the model, and starting values of 2 and 4 for σ_u^2 and σ_e^2 , respectively, in the random part of the model. For illustrative purposes, we also dispense with the burnin, allowing us to subsequently view the first 500 iterations, as these will be most influenced by the starting values:

```
R> OurStartingValues <- list(FP.b = c(-2, 5), RP.b = c(2, 4))
R> startvalMCMC <- runMLwiN(F3, data = tutorial,
+   estoptions = list(EstM = 1, startval = OurStartingValues,
+   mcmcMeth = list(burnin = 0, iterations = 500)))
```

```
R> trajectories(startvalMCMC["chains"], Range = c(1, 500))
```

Here we can see, in Figure 6, the chains quickly moving from the starting values we have given them to values closer to those we would have received from an initial IGLS fit.

Finally, it is also a simple matter to change the random number seed from the default of 1 (e.g., `estoptions = list(EstM = 1, mcmcMeth = list(seed = 2))`).

8. Adding a random slope/coefficient

In the modelling so far we have assumed that only the intercept term in our models varies across clusters. We have already included `standlrt` in the fixed part of the model to allow for a fixed slope effect; if we also include it to the left of `| school` in the random part of the model, we will additionally allow the coefficient of our predictor to randomly vary across level 2 units, thus specifying Equation 4.

$$\begin{aligned} \text{normexam}_{ij} &= \beta_0 + \beta_1 \text{standlrt}_{ij} + u_{0j} + u_{1j} \text{standlrt}_{ij} + e_{ij} \\ \begin{pmatrix} u_{0j} \\ u_{1j} \end{pmatrix} &\sim N \left\{ \begin{pmatrix} 0 \\ 0 \end{pmatrix}, \begin{pmatrix} \sigma_{u0}^2 & \\ & \sigma_{u1}^2 \end{pmatrix} \right\} \\ e_{ij} &\sim N(0, \sigma_e^2) \end{aligned} \quad (4)$$

```
R> F4 <- normexam ~ 1 + standlrt + (1 + standlrt | school) + (1 | student)
R> (standlrtRS_MCMC <- runMLwiN(
+   Formula = F4, data = tutorial,
+   estoptions = list(EstM = 1, resi.store.levs = 2)))
```

```
-----
MLwiN (version: 2.36) multilevel model (Normal)
      N min      mean max N_complete min_complete mean_complete max_complete
school 65    2 62.44615 198           65           2          62.44615          198
Estimation algorithm: MCMC      Elapsed time : 2.12s
Number of obs:4059 (from total 4059) Number of iter:5000 Chains:1 Burn-in:500
Bayesian Deviance Information Criterion (DIC)
Dbar      D(thetabar)    pD      DIC
9122.986   9031.321     91.666   9214.652
-----
```

The model formula:

```
normexam ~ 1 + standlrt + (1 + standlrt | school) + (1 | student)
Level 2: school      Level 1: student
-----
```

The fixed part estimates:

	Coef.	Std. Err.	z	Pr(> z)	[95% Cred. Interval]	ESS
Intercept	-0.00569	0.03923	-0.15	0.8847	-0.08045 0.07369	233
standlrt	0.55827	0.02047	27.27	9.883e-164 ***	0.51762 0.59858	769

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

The random part estimates at the school level:

	Coef.	Std. Err.	[95% Cred.	Interval]	ESS
var_Intercept	0.09600	0.01985	0.06396	0.13998	3117
cov_Intercept_standlrt	0.01941	0.00723	0.00681	0.03476	1809
var_standlrt	0.01547	0.00450	0.00810	0.02561	1063

 The random part estimates at the student level:

	Coef.	Std. Err.	[95% Cred.	Interval]	ESS
var_Intercept	0.55427	0.01257	0.52984	0.57961	5189

Here we see that including such a random slope increases the effective number of parameters (pD), but the reduction in DIC indicates that, despite this increase in complexity, it is a better model:

```
R> ComparingModels <- rbind(standlrtMCMC["BDIC"], standlrRS_MCMC["BDIC"])
R> rownames(ComparingModels) <- c("Random intercept", "Random slope")
R> ComparingModels
```

	Dbar	D(thetabar)	pD	DIC
Random intercept	9209.146	9149.164	59.98223	9269.128
Random slope	9122.986	9031.321	91.66588	9214.652

We included `resi.store.levs = 2` in our list of `estoptions`, above, to illustrate **R2MLwiN**'s `predLines` function; this draws predicted lines against an explanatory variable for each group at a higher level, calculating the median and quantiles in doing so (note that `predLines` uses a lot of contiguous memory, so we recommend using the 64-bit version of R to mitigate against any problems this may cause). For example, the script below produces a plot of all 65 school lines, plus their intervals (see Figure 7). As well as specifying the model object, we also provide the name of the predictor (`xname`), the level (`lev`), and the probabilities (`probs`) used to calculate the lower and upper quantiles from which the error bars are plotted:

```
R> predLines(standlrRS_MCMC, xname = "standlrt", lev = 2,
+   probs = c(0.025, 0.975), legend = FALSE)
```

By not declaring otherwise, we have accepted the default of `selected = NULL` and have thus produced quite a busy plot, although a general fanning-out pattern is apparent, reflecting the positive intercept/slope covariance at the school level (`cov_Intercept_standlrt`, in the model output above). To better distinguish individual schools, however, we can instead choose to select just a few (requesting a `legend` whilst doing so; see Figure 8):

```
R> predLines(standlrRS_MCMC, xname = "standlrt", lev = 2,
+   selected = c(30, 44, 53, 59), probs = c(0.025, 0.975), legend = TRUE)
```

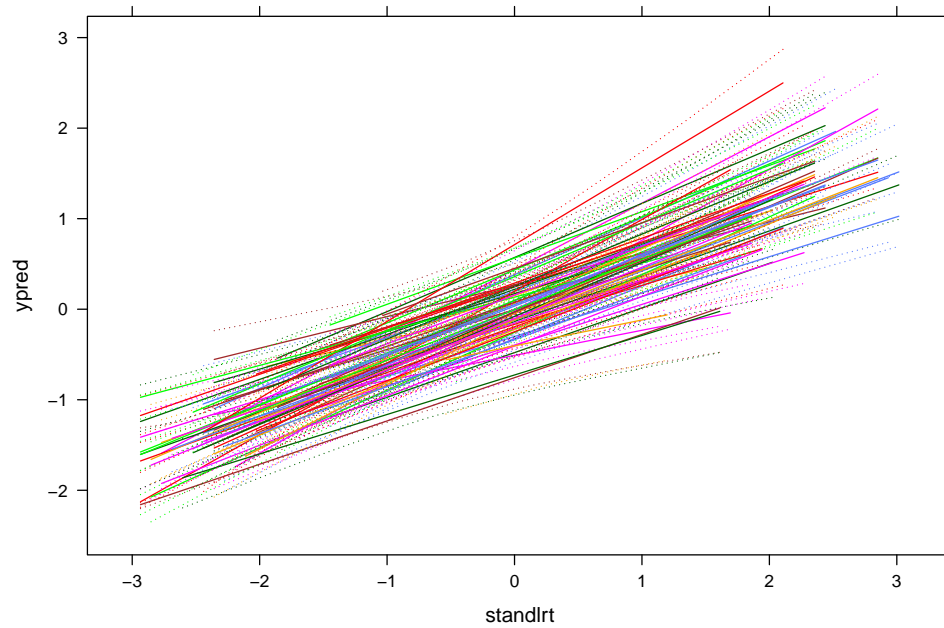


Figure 7: Predicted lines (with 95% credible intervals) for each of the 65 schools in the **tutorial** dataset.

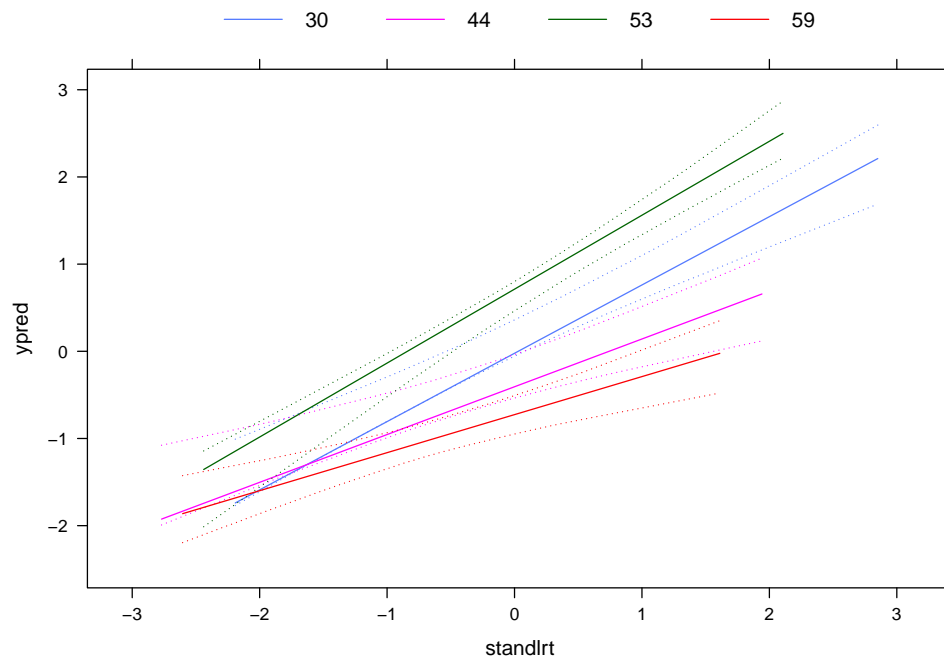


Figure 8: Predicted lines (with 95% credible intervals) for four schools in the **tutorial** dataset.

9. Modeling complex level 1 variance

Another important aspect of multilevel models is their ability to model complex level 1 variance – i.e., residual heteroscedasticity. Whilst it is not currently possible to fit complex level 1 variance models in R packages such as `lme4` (Bates *et al.* 2015), it is possible to do so via MLwiN (using both IGLS and MCMC, although here we focus on the latter). For example, taking the model we have just fitted above, we can allow for the possibility that variability in `normexam` within-schools (i.e., around their fitted regression lines) may also depend on `standlrt` (see Browne *et al.* 2002, for an algorithm to fit this extension).

When fitting this model below, note that we have chosen to include `mcmcMeth = list(lclo = 1)` in our list of `estoptions` so that MLwiN will fit the log of the precision (1/variance) at level 1 as a function of the predictors; since this can take any value on the real line, we are free of the restrictions on prior distributions which result from fitting the variance as a linear function of the predictors (see Spiegelhalter *et al.* 2000):

```
R> F5 <- normexam ~ 1 + standlrt + (1 + standlrt | school) +
+   (1 + standlrt | student)
R> standlrtC1V_MCMC <- runMLwiN(
+   Formula = F5, data = tutorial,
+   estoptions = (list(EstM = 1, mcmcMeth = list(lclo = 1))))
```

Having fitted this model, we can then explore how the variance changes both within and between schools for different values of the intake score, `standlrt`, for example in a plot (Figure 9):

```
R> l2varfn <- standlrtC1V_MCMC["RP"][["RP2_var_Intercept"]] +
+   2 * standlrtC1V_MCMC["RP"][["RP2_cov_Intercept_standlrt"]] *
+   tutorial[["standlrt"]] + standlrtC1V_MCMC["RP"][["RP2_var_standlrt"]] *
+   tutorial[["standlrt"]]^2
R> l1varfn <- 1 / exp(standlrtC1V_MCMC["RP"][["RP1_var_Intercept"]] +
+   2 * standlrtC1V_MCMC["RP"][["RP1_cov_Intercept_standlrt"]] *
+   tutorial[["standlrt"]] + standlrtC1V_MCMC["RP"][["RP1_var_standlrt"]] *
+   tutorial[["standlrt"]]^2)
R> plot(sort(tutorial[["standlrt"]]),
+   l2varfn[order(tutorial[["standlrt"]])], xlab = "standlrt",
+   ylab = "variance", ylim = c(0,.7), type = "l")
R> lines(sort(tutorial[["standlrt"]]),
+   l1varfn[order(tutorial[["standlrt"]])], lty = "longdash")
R> abline(v = 0, lty = "dotted")
R> legend("bottomright", legend = c("between-student", "between-school"),
+   lty = c("longdash", "solid"))
```

Figure 9 indicates that the between-school variation generally increases as `standlrt` increases, whereas the opposite is true for the between-student (within-school) variation.

Finally, the following sample of three trajectory plots, plotting only the last 500 iterations (see Figure 10), illustrate that MLwiN has recognized that a different sampling method – Metropolis Hastings (MH), with its characteristic block-like appearance – is necessary to update the level 1 variance parameters, with Gibbs sampling updating the other parameters.

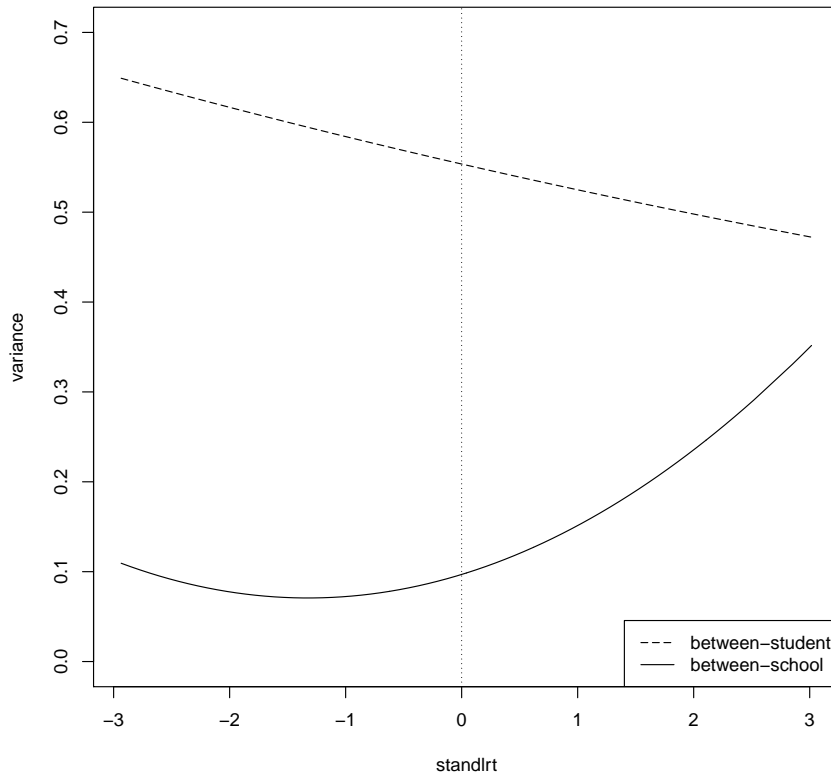


Figure 9: A plot of the variance function from a random slopes model allowing for heterogeneity at level 1.

```
R> trajectories(standlrtC1V_MCMC["chains"][, "FP_Intercept",
+ drop = FALSE], Range = c(4501, 5000))
R> trajectories(standlrtC1V_MCMC["chains"][, "RP2_var_Intercept",
+ drop = FALSE], Range = c(4501, 5000))
R> trajectories(standlrtC1V_MCMC["chains"][, "RP1_var_Intercept",
+ drop = FALSE], Range = c(4501, 5000))
```

10. Fitting a 2-level binary response model via MCMC

We will now move on from continuous response models to consider some of the other response distribution types supported by **R2MLwiN**. As Table 2 illustrates, many of these, including the "Binomial" distribution we explore in the next example, can be fitted using IGLS or MCMC estimation; here, though, we will stay with MCMC in order to illustrate alternative MCMC methodology implemented in MLwiN, and also interoperability with BUGS, in the following sections.

Our example dataset is a sub-sample of the 1989 Bangladesh Fertility Survey (Huq and Cleland 1990), provided as `bang1` in both MLwiN and **R2MLwiN** (see Table 3 for a list of the variables to be modeled).

Below we construct a logit model to investigate whether the use of contraceptives at the time

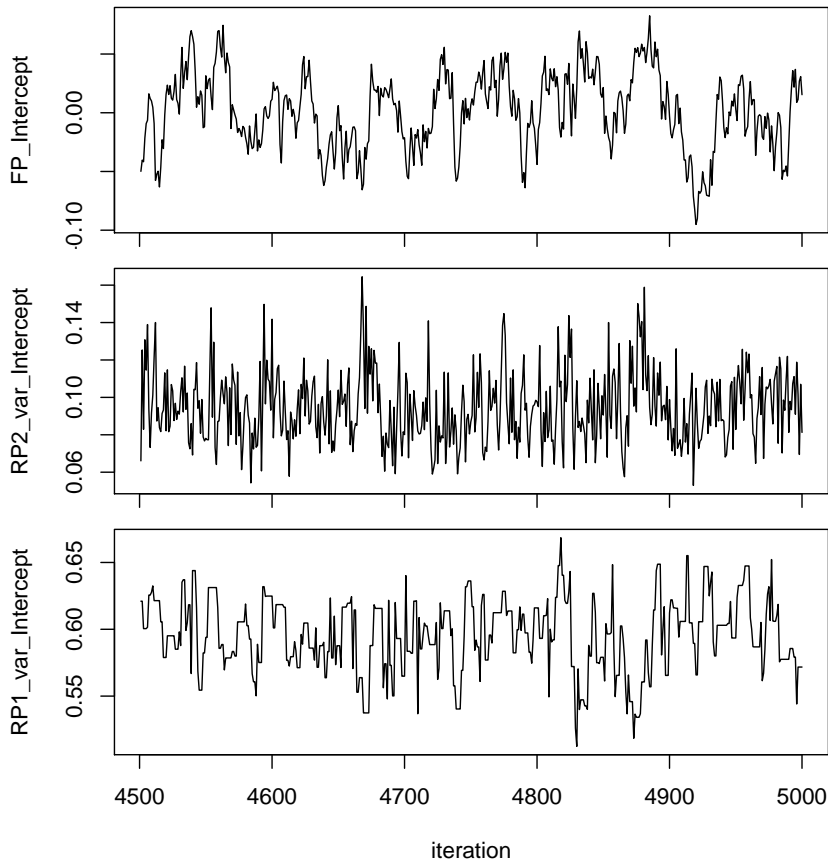


Figure 10: Modeling complex level 1 variance: chain trajectories of (from top) β_0 , σ_{u0}^2 , and σ_{e0}^2 .

Variable	Description
woman	Identifying code for each woman.
district	Identifying code for each district.
use	Contraceptive use status at time of survey; levels are <code>Not_using</code> and <code>Using</code> .
lc	Number of living children at time of survey; levels correspond to none (<code>None</code>), one (<code>One_child</code>), two (<code>Two_children</code>), or three or more children (<code>Three_plus</code>).
age	Age of woman at time of survey (in years), centered on the sample mean of 30 years.
urban	Type of region of residence; levels are <code>Rural</code> and <code>Urban</code> .

Table 3: Variables in the `bang1` dataset, as modeled in the worked example.

of the survey is predicted by the `age` of the respondent, their number of children living at the time of the survey (`lc`), and the type of region in which they reside (`urban`); furthermore, we will allow the coefficient of the latter predictor to randomly-vary at the `district`-level to explore whether variability between `Urban` areas within districts differs from that between

Rural areas within districts, producing the random coefficient model in Equation 5.

$$\begin{aligned} \text{use}_{ij} &\sim \text{Bernoulli}(\pi_{ij}) \\ \text{logit}(\pi_{ij}) &= \beta_0 + \beta_1 \text{age}_{ij} + \beta_2 \text{lcOne_child}_{ij} + \beta_3 \text{lcTwo_children}_{ij} + \beta_4 \text{lcThree_plus}_{ij} \\ &\quad + \beta_5 \text{urban}_{ij} + u_{0j} + u_{5j} \text{urban}_{ij} \\ \begin{pmatrix} u_{0j} \\ u_{5j} \end{pmatrix} &\sim \text{N} \left\{ \begin{pmatrix} 0 \\ 0 \end{pmatrix}, \begin{pmatrix} \sigma_{u0}^2 & \\ & \sigma_{u5}^2 \end{pmatrix} \right\} \end{aligned} \tag{5}$$

We can fit this, via **R2MLwiN**, as follows:

```
R> data("bang1")
R> F6 = logit(use) ~ 1 + age + lc + urban + (1 + urban | district)
R> binomialMCMC <- runMLwiN(Formula = F6, D = "Binomial", data = bang1,
+   estoptions = list(EstM = 1))
R> trajectories(binomialMCMC["chains"][, "FP_Intercept", drop = FALSE])
```

As discussed in Table 2, the terms in the parentheses immediately following `logit` are of the form **response variable**, **denominator**; specifying the denominator is actually optional, and if not specified will take the form of a constant of ones, which is appropriate here. Since MLwiN requires binary response variables to be of the form 0/1, **R2MLwiN** transforms it into that format having detected it is indeed binary, and thus we are modeling a Bernoulli distribution. `lc` is a **factor** variable and so dummy variables will be added as predictors with `None` (see `min(levels(bang1$lc))`) as the reference category (the `relevel` function, part of the **stats** package, can be used to assign an alternative reference category, by re-ordering the levels of a factor prior to an `runMLwiN` call).

We have requested, on the last line, a plot of the chain trajectory for the intercept in the fixed part of the model. As the resulting plot (Figure 11) indicates, mixing here looks relatively poor, and so it may be worthwhile examining some of the alternative MCMC methods implemented in MLwiN; we review these, and apply one to the current example, in the next section.

11. Alternative MCMC methods implemented in MLwiN

MLwiN's MCMC engine features a number of methods which can improve the efficiency of MCMC estimation (see Browne 2012, chapters 21-25). Table 4 provides a summary of available methods, one of which – orthogonal parameterization – we explore in the example below, before moving on to discuss further options via MLwiN's interoperability with BUGS.

11.1. An example using orthogonal parameterization to improve mixing

Given that we encountered poor mixing in our last model fit, above, we will examine whether orthogonal parameterization may help; this replaces a set of fixed effect predictors with an alternative group of predictors that span the same parameter space, but are orthogonal, and is particularly useful for instances such as this, where we are modeling a non-normal distribution (since fixed effects in normal models are generally updated in a block).

Here, then, we redefine `estoptions`, toggling `orth` to 1 in the list of `mcmcOptions`, and run the model again:

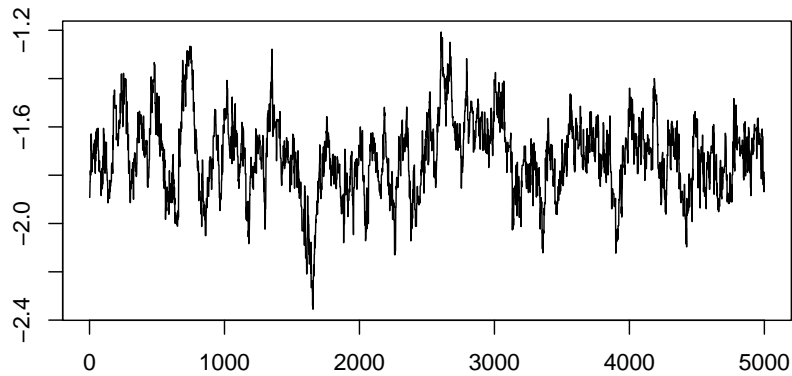


Figure 11: Chain trajectory of β_0 from a binomial response model, without orthogonal parametrization.

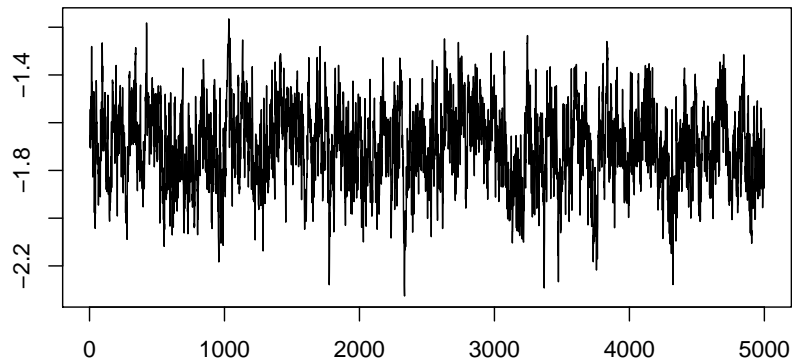


Figure 12: Chain trajectory of β_0 from a binomial response model, with orthogonal parametrization.

```
R> (OrthogbinomialMCMC <- runMLwiN(Formula = F6, D = "Binomial",
+   data = bang1, estoptions = list(EstM = 1, mcmcOptions = list(orth = 1))))
```

```
-----
MLwiN (version: 2.36) multilevel model (Binomial)
      N min  mean max N_complete min_complete mean_complete max_complete
district 60  2 32.23333 118         60           2         32.23333         118
Estimation algorithm: MCMC      Elapsed time : 15.06s
Number of obs:1934 (from total 1934) Number of iter:5000 Chains:1 Burn-in:500
Bayesian Deviance Information Criterion (DIC)
Dbar      D(thetabar)    pD      DIC
2329.877  2273.176    56.702  2386.579
-----
```

```
The model formula:
logit(use) ~ 1 + age + lc + urban + (1 + urban | district)
Level 2: district      Level 1: l1id
-----
```

The fixed part estimates:

Method	As currently implemented in MLwiN	As specified in <code>mcmcOptions</code>
Structured MCMC (e.g., Sargent et al. 2009)	2-level nested normal response models, with no complex level 1 variation, only	<code>smcm = 1</code>
Structured MVN framework (e.g., Browne et al. 2009a)	2-level variance components models only	<code>smvn = 1</code>
Othogonal fixed effect vectors (e.g., Browne et al. 2009b)	All models	<code>orth = 1</code>
Parameter expansion (e.g., Liu and Wu 1999)	All models	<code>paex = c(<L>, 1)</code>
Hierarchical centring (e.g., Gelfand et al. 1995)	All models	<code>hcen = <L></code>

Table 4: A summary of some alternative methods for MCMC estimation available via the **R2MLwiN** package. `<L>` is an integer specifying the level at which parameter expansion / hierarchical centering (respectively) is to occur. Taking Structured MCMC as an example, these methods are implemented as follows: `estoptions = list(EstM = 1, mcmcOptions = list(smcm = 1), ...)`.

```

              Coef. Std. Err.      z    Pr(>|z|)    [95% Cred. Interval]  ESS
Intercept    -1.70164  0.16702 -10.19 2.237e-24 ***  -2.03503  -1.38148  246
age          -0.02645  0.00798  -3.31 0.0009172 ***  -0.04191  -0.01117 1063
lcOne_child   1.12627  0.16211   6.95 3.716e-12 ***   0.80576   1.45134  923
lcTwo_children 1.34947  0.17658   7.64 2.133e-14 ***   1.00799   1.69571  921
lcThree_plus  1.34643  0.18528   7.27 3.676e-13 ***   0.97975   1.70612  872
urbanUrban    0.81100  0.19211   4.22 2.426e-05 ***   0.45403   1.22458  151
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

The random part estimates at the district level:

```

              Coef.  Std. Err.  [95% Cred.  Interval]  ESS
var_Intercept      0.41681    0.13544    0.21169    0.73256   223
cov_Intercept_urbanUrban -0.41923    0.17235   -0.83135   -0.16282   139
var_urbanUrban      0.70042    0.29966    0.28379    1.43387   102

```

The random part estimates at the llid level:

```

              Coef.  Std. Err.  [95% Cred.  Interval]  ESS
var_bcons_1  1.00000    1e-05    1.00000    1.00000   5000

```

```
R> trajectories(OrthogbinomialMCMC["chains"][, "FP_Intercept", drop = FALSE])
```

As such, we see that the mixing is considerably better (Figure 12), with the ESS considerably-improved for the fixed effects (see comparison table in the following section). The model indicates that the probability of contraceptive use decreases with `age`, but increases with the number of living children, and is greater in `Urban` areas; furthermore, the variance in contra-

ceptive use differs between Rural areas (estimated as 0.417), and Urban areas (estimated as $0.417 - 2 \times 0.419 + 0.7 = 0.28$).

12. Using R2MLwiN to write BUGS code

As well as using its own MCMC estimation engine, MLwiN can write code to fit models in **WinBUGS** (Lunn *et al.* 2000) and **OpenBUGS** (Lunn *et al.* 2009), with the inclusion of its own IGLS estimates as starting values. With the aid of the **rbugs** package (Yan and Prates 2013), the user can employ a single `runMLwiN` function call to obtain starting values from an IGLS run in MLwiN, automatically generate the necessary BUGS model code, initial values, data files, and script, and then fit the model in BUGS.

As demonstrated in the script below, **WinBUGS** and **OpenBUGS** are called via the `runMLwiN` argument `BUGO`; this is a vector where `n.chains` corresponds to the number of chains, `debug` determines whether BUGS stays open following completion of the model run, `seed` sets the random number seed, `bugs` specifies the path, and `OpenBUGS` can toggle between `FALSE` for **WinBUGS**, and `TRUE` for **OpenBUGS**. Here we fit the same model as in the example above (the `burnin`, `iterations` and `thinning` we specify are the defaults, we simply make them explicit here for the purposes of this example); by specifying `show.file = TRUE` in the list of `estoptions`, the BUGS model is returned (see supplementary materials):

```
R> WinBUGS <- "C:/WinBUGS14/WinBUGS14.exe"
R> BUGSmodel = runMLwiN(Formula = F6, D = "Binomial",
+   data = bang1, estoptions = list(EstM = 1,
+   mcmcMeth = list(burnin = 500, iterations = 5000, thinning = 1)),
+   BUGO = c(n.chains = 1, debug = FALSE, seed = 1,
+   bugs = WinBUGS, OpenBugs = FALSE))
R> summary(BUGSmodel)
```

```
Iterations = 1:5000
Thinning interval = 1
Number of chains = 1
Sample size per chain = 5000
```

1. Empirical mean and standard deviation for each variable, plus standard error of the mean:

	Mean	SD	Naive SE	Time-series SE
beta[1]	-1.721e+00	0.161037	0.0022774	0.0097350
beta[2]	-2.675e-02	0.007946	0.0001124	0.0002641
beta[3]	1.140e+00	0.158146	0.0022365	0.0053347
beta[4]	1.366e+00	0.179230	0.0025347	0.0068177
beta[5]	1.365e+00	0.181054	0.0025605	0.0087925
beta[6]	8.136e-01	0.171615	0.0024270	0.0081912
deviance	2.329e+03	14.099908	0.1994028	0.5284579
sigma2.u2[1,1]	4.216e-01	0.138640	0.0019607	0.0063190
sigma2.u2[1,2]	-4.350e-01	0.187684	0.0026542	0.0098587

```

sigma2.u2[2,1] -4.350e-01  0.187684  0.0026542      0.0098587
sigma2.u2[2,2]  7.283e-01  0.341042  0.0048231      0.0199511
u2[1,1]        -9.429e-01  0.334778  0.0047345      0.0118369
u2[1,2]        3.969e-01  0.417123  0.0058990      0.0149241
...

```

On finishing the model run, the model estimates are returned (including the quantiles, not displayed in the truncated output above), but this time as a `summary` of an `mcmc.list` (see `coda` (Plummer *et al.* 2006)).

Running this on our machine took 140 seconds in **WinBUGS**, which uses (by default) a multivariate Metropolis update of the fixed effects (Gameran 1997), whilst it took 16 seconds to run the model for the same length of chain in MLwiN (both via **R2MLwiN**), which uses a single site Metropolis sampler. As the table below indicates, however, the effective sample sizes (ESSs) from the run in BUGS are considerably larger than those from the run (without orthogonal parameterization) in MLwiN (although there is considerable improvement when we opt for orthogonal parameterisation in MLwiN, at least for the fixed effects; this took 15 seconds to run on our machine).

```

R> cc = cbind(binomialMCMC["FP"], OrthogbinomialMCMC["FP"])
R> dd = cbind(head(binomialMCMC["RP"], -1),
+   head(OrthogbinomialMCMC["RP"], -1))
R> ESS.binMCMC = effectiveSize(binomialMCMC["chains"][,2:10])
R> ESS.orthogMCMC = effectiveSize(OrthogbinomialMCMC["chains"][,2:10])
R> ESStable = round(rbind(cc, dd), 3)
R> BUGSlist <- c(1:6, 8, 9, 11)
R> BUGS.Coeff <- round(summary(BUGSmodel)$statistics[BUGSlist, 1], 3)
R> BUGS.ESS <- as.data.frame(effectiveSize(BUGSmodel))
R> ESStable = cbind(ESStable[, 1], round(ESS.binMCMC), ESStable[, 2],
+   round(ESS.orthogMCMC), BUGS.Coeff, round(BUGS.ESS[BUGSlist, ]))
R> colnames(ESStable) = c("A)Coeff.", "A)ESS", "B)Coeff.",
+   "B)ESS", "C)Coeff.", "C)ESS")
R> cat("NB: A = MLwiN(non-orthog.), B = MLwiN(orthog.), C = WinBUGS\n");
+   ESStable

```

NB: A = MLwiN(non-orthog.), B = MLwiN(orthog.), C = WinBUGS

	A)Coeff.	A)ESS	B)Coeff.	B)ESS	C)Coeff.	C)ESS
FP_Intercept	-1.724	61	-1.702	246	-1.721	274
FP_age	-0.027	203	-0.026	1063	-0.027	905
FP_lcOne_child	1.157	229	1.126	923	1.140	879
FP_lcTwo_children	1.376	186	1.349	921	1.366	691
FP_lcThree_plus	1.384	102	1.346	872	1.365	424
FP_urbanUrban	0.805	110	0.811	151	0.814	439
RP2_var_Intercept	0.418	206	0.417	223	0.422	481
RP2_cov_Intercept_urbanUrban	-0.432	127	-0.419	139	-0.435	362
RP2_var_urbanUrban	0.738	142	0.700	102	0.728	292

13. Modeling a cross-classified data structure

So far the multilevel models we have examined have been strictly hierarchical, but researchers may well encounter data structures which are cross-classified (i.e., crossed random effects), exhibit multiple membership, or indeed a combination of these; **R2MLwiN** allows users to fit such models via MCMC estimation.

In the following example we use an educational dataset from Fife in Scotland (available as the sample dataset `xc` in **MLwiN** and **R2MLwiN**); see Table 5 for a description of the variables we will model.

We wish to model students' examination attainment, whilst controlling for clustering due to their current (secondary) school (`sid`), but also controlling for clustering due to their (earlier) primary school too (`pid`); since not all students who attended a given primary school all subsequently attended the same secondary school, the former are not nested within the latter, and so we have a cross-classified structure; see Equation 6, which uses the classification notation as per Browne *et al.* (2001).

$$\begin{aligned} \text{attain}_i &= \beta_0 + u_{\text{sid}(i)}^{(2)} + u_{\text{pid}(i)}^{(3)} + e_i \\ u_{\text{sid}(i)}^{(2)} &\sim N(0, \sigma_{u(2)}^2) \\ u_{\text{pid}(i)}^{(3)} &\sim N(0, \sigma_{u(3)}^2) \\ e_i &\sim N(0, \sigma_e^2) \end{aligned} \tag{6}$$

As indicated below, we need to specify that the data structure is cross-classified by declaring `xc = TRUE` within the list of `estoptions`:

```
R> data("xc")
R> F7 = attain ~ 1 + (1 | sid) + (1 | pid) + (1 | pupil)
R> (XCModel <- runMLwiN(Formula = F7, data = xc,
+   estoptions = list(xc = TRUE, EstM = 1)))
```

```
-----
MLwiN (version: 2.36) multilevel model (Normal)
      N min      mean max N_complete min_complete mean_complete max_complete
sid  19  92 180.78947 290          19           92      180.78947          290
pid 148   1  23.20946  72          148           1       23.20946           72
Estimation algorithm: MCMC      Cross-classified      Elapsed time : 2.12s
Number of obs:3435 (from total 3435) Number of iter:5000 Chains:1 Burn-in:500
```

Variable	Description
<code>attain</code>	Attainment score of pupils at age 16.
<code>pid</code>	Primary school identifying code.
<code>sid</code>	Secondary school identifying code.
<code>pupil</code>	Pupil identifying code.

Table 5: Variables in the `xc` dataset, as modeled in the worked example.

Bayesian Deviance Information Criterion (DIC)

Dbar	D(thetabar)	pD	DIC
16940.564	16833.400	107.164	17047.729

The model formula:

```
attain ~ 1 + (1 | sid) + (1 | pid) + (1 | pupil)
Level 3: sid      Level 2: pid      Level 1: pupil
```

The fixed part estimates:

	Coef.	Std. Err.	z	Pr(> z)		[95% Cred. Interval]	ESS
Intercept	5.50404	0.18959	29.03	2.69e-185 ***		5.10783 5.86623	225

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

The random part estimates at the sid level:

	Coef.	Std. Err.	[95% Cred. Interval]	ESS
var_Intercept	0.41403	0.20965	0.14194 0.92900	1042

The random part estimates at the pid level:

	Coef.	Std. Err.	[95% Cred. Interval]	ESS
var_Intercept	1.15258	0.21475	0.79583 1.62561	1319

The random part estimates at the pupil level:

	Coef.	Std. Err.	[95% Cred. Interval]	ESS
var_Intercept	8.11992	0.20175	7.73324 8.52689	4657

As such we see (below) that 11.9% of the variation in exam attainment at age 16 is attributable to primary schools with less, 4.3%, attributable to secondary schools, possibly because the latter are larger, more socially-heterogeneous, institutions (e.g., [Goldstein 2010](#)):

```
R> PercentExplainedBy <- cbind(
+   XCMoel["RP"][["RP3_var_Intercept"]] /
+   (XCMoel["RP"][["RP1_var_Intercept"]] +
+   XCMoel["RP"][["RP2_var_Intercept"]] +
+   XCMoel["RP"][["RP3_var_Intercept"]]) * 100,
+   XCMoel["RP"][["RP2_var_Intercept"]] /
+   (XCMoel["RP"][["RP1_var_Intercept"]] +
+   XCMoel["RP"][["RP2_var_Intercept"]] +
+   XCMoel["RP"][["RP3_var_Intercept"]]) * 100)
R> colnames(PercentExplainedBy) <- c("sid", "pid")
R> PercentExplainedBy
```

	sid	pid
[1,]	4.274312	11.89881

14. Modeling a multiple membership data structure

Multiple membership models arise when we wish to control for clustering at a given ‘level’ or classification, but have lower-level units which belong to more than one group at that higher level. For example, if we wished to model employees’ earnings over the past financial year, we might want to control for non-independence due to the companies which employed them during this time. If the employees were employed by more than one company, however (i.e., they were a ‘member’ of more than one group at this higher level), it would be judicious to control for the effects of all of them.

Below we fit such a model, using the `wage1` sample dataset (available in both `MLwiN` and `R2MLwiN`); see Table 6 for a description of the variables we will model, and Equation 7 for the model itself.

$$\begin{aligned} \text{logearn}_i &= \beta_0 + \beta_1 \text{age_40}_i + \beta_2 \text{numjobs}_i + \sum_{j \in \text{company}(i)} w_{i,j}^{(2)} u_j^{(2)} + e_i \\ i &= 1, \dots, N \\ \text{company}(i) &\subset (1, \dots, J) \\ u_j^{(2)} &\sim N(0, \sigma_{u^{(2)}}^2) \\ e_i &\sim N(0, \sigma_e^2) \end{aligned} \tag{7}$$

You can see, below, that the argument `mm`, specifying the structure of the multiple membership model, has been included in the list of `estoptions` (NB since `mm` is non-NULL, `xc` defaults to `TRUE`). The argument `mm` can be a list of variable names, a list of vectors, or a matrix (e.g., see `?df2matrix`). Here we employ variable names, which need to be assigned as lists to `mmvar`, specifying the classification units, and `weights`, specifying the weights. In the

Variable	Description
<code>id</code>	Identifying code for each office worker.
<code>company</code>	Identifying code for first company worked for over the last 12 months.
<code>company2</code>	If worked for > 1 company over the last 12 months, identifying code for second company.
<code>company3</code>	If worked for > 2 company over the last 12 months, identifying code for third company.
<code>company4</code>	If worked for > 3 company over the last 12 months, identifying code for fourth company.
<code>logearn</code>	Workers’ (natural) log-transformed earnings over the last financial year.
<code>numjobs</code>	The number of companies worked for over the last 12 months.
<code>weight1</code>	Proportion of time worked for employer listed in <code>company</code> .
<code>weight2</code>	Proportion of time worked for employer listed in <code>company2</code> .
<code>weight3</code>	Proportion of time worked for employer listed in <code>company3</code> .
<code>weight4</code>	Proportion of time worked for employer listed in <code>company4</code> .
<code>age_40</code>	Age of workers, centered on 40 years.

Table 6: Variables in the `wage1` dataset, as modeled in the worked example.

variables comprising the `mmvar` list, the same company, e.g., ACME Ltd, has the same value, e.g., 8, throughout. The `weights` specify the employee-level weighting given to each company they worked for; here we have chosen the proportion of time an employee has spent with a particular company. Each element of the list corresponds to a level (classification) of the model, in descending order, so the final NA indicates that there is not a multiple membership classification at level 1.

```
R> data("wage1")
R> F8 = logearn ~ 1 + age_40 + numjobs + (1 | company) + (1 | id)
R> OurMultiMemb <- list(list(
+   mmvar = list("company", "company2", "company3", "company4"),
+   weights = list("weight1", "weight2", "weight3", "weight4")), NA)
R> (MMembModel <- runMLwiN(Formula = F8, data = wage1,
+   estoptions = list(EstM = 1, drop.data = FALSE, mm = OurMultiMemb)))
```

```
MLwiN (version: 2.36) multilevel model (Normal)
      N min   mean max N_complete min_complete mean_complete max_complete
company 141  2 21.43262 49      141           2      21.43262      49
Estimation algorithm: MCMC      Cross-classified      Elapsed time : 1.46s
Number of obs:3022 (from total 3022) Number of iter:5000 Chains:1 Burn-in:500
Bayesian Deviance Information Criterion (DIC)
Dbar      D(thetabar)      pD      DIC
4625.279  4514.715      110.564  4735.843
```

```
The model formula:
logearn ~ 1 + age_40 + numjobs + (1 | company) + (1 | id)
Level 2: company      Level 1: id
```

```
The fixed part estimates:
      Coef. Std. Err.      z Pr(>|z|)      [95% Cred. Interval]      ESS
Intercept  3.04872  0.03534  86.26      0 ***      2.98024  3.11807  851
age_40     0.01273  0.00098  13.06 5.924e-39 ***      0.01084  0.01460  4273
numjobs    -0.11190  0.02223  -5.03 4.803e-07 ***      -0.15446 -0.06892  4594
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
The random part estimates at the company level:
      Coef. Std. Err.      [95% Cred. Interval]      ESS
var_Intercept  0.05793  0.00916      0.04225  0.07808  2196
```

```
The random part estimates at the id level:
      Coef. Std. Err.      [95% Cred. Interval]      ESS
var_Intercept  0.27064  0.00711      0.25717  0.28472  4614
```

```
R> MMembModel[["RP"]][["RP2_var_Intercept"]] /
+   (MMembModel[["RP"]][["RP2_var_Intercept"]] +
+   MMembModel[["RP"]][["RP1_var_Intercept"]]) * 100
```

[1] 17.63173

The model indicates (as calculated at the bottom) that `company` accounted for 17.6% of the variance in (log-transformed) earnings, having controlled for age and number of jobs. Looking at the fixed effects, earnings on average increase with age, but those employees working for more companies over the past 12 months earn on average less; it would be interesting to investigate whether these effects persist once other variables available in the dataset are taken into account, such as sex of the employee, and whether they work part-time or not, but we will leave this model as it is, and move on to our next example.

15. Multivariate response models

Here we illustrate fitting a multivariate response model. In this example dataset (see [Browne 2012](#)), taken from the larger junior school project (JSP) dataset ([Mortimore et al. 1988](#)), we have two responses (see [Table 7](#)): `english` – an English test score marked out of 100, and `behaviour` – a binary behaviour score taken in the same year. We’re interested in the correlation between English test performance and behaviour, and the effect of predictors – such as `sex`, and an earlier test score (`ravens`) – on both of them, and also the effect of an earlier English fluency indicator (`fluent`) on the `english` test (i.e., not `behaviour`, since preliminary investigation revealed no correlation between the two); see [Equation 8](#).

Note that the MCMC engine in MLwiN can fit mixed response multivariate models, as well as normal response models, but only if they consist of a mixture of normal responses and binary responses (with a probit, rather than logit, link), as in this case. In practice latent normal variables are constructed for each binary response with the value of the binary response governing the sign of the latent variable as illustrated below:

$$\begin{aligned}
 \text{english}_{ij} &= \beta_0 + \beta_1 \text{sex}_{ij} + \beta_2 \text{ravens}_{ij} + \beta_3 \text{fluent}_{ij} + u_{0j} + e_{0ij} \\
 \text{behaviour}_{ij}^* &= \beta_4 + \beta_5 \text{sex}_{ij} + \beta_6 \text{ravens}_{ij} + u_{4j} + e_{4ij} \\
 \text{where } \text{behaviour}_{ij}^* &\geq 0 \text{ if } \text{behaviour}_{ij} = 1 \\
 \text{and } \text{behaviour}_{ij}^* &< 0 \text{ if } \text{behaviour}_{ij} = 0
 \end{aligned} \tag{8}$$

$$\begin{aligned}
 \begin{pmatrix} u_{0j} \\ u_{4j} \end{pmatrix} &\sim N \left\{ \begin{pmatrix} 0 \\ 0 \end{pmatrix}, \begin{pmatrix} \sigma_{u0}^2 & \\ \sigma_{u04} & \sigma_{u4}^2 \end{pmatrix} \right\} \\
 \begin{pmatrix} e_{0ij} \\ e_{4ij} \end{pmatrix} &\sim N \left\{ \begin{pmatrix} 0 \\ 0 \end{pmatrix}, \begin{pmatrix} \sigma_{e0}^2 & \\ \sigma_{e04} & 1 \end{pmatrix} \right\}
 \end{aligned}$$

```

R> data("jspmix1")
R> F9 = c(english, probit(behaviour)) ~ 1 + sex + ravens + fluent[1] +
+   (1 | school) + (1[1] | id)
R> (MixedRespMCMC <- runMLwiN(Formula = F9, D = c("Mixed", "Normal",
+   "Binomial"), data = jspmix1, estoptions = list(EstM = 1,
+   mcmcMeth = list(fixM = 1, residM = 1, Lev1VarM = 2))))

```

Here then we have two terms to the left of the tilde (\sim) in the model formula, corresponding to the two responses (their order matters, insofar as we use it to distinguish them when referring to the response variables in other parts of our model specification). Firstly we have `english`,

Variable	Description
<code>school</code>	Identifying code for school.
<code>id</code>	Identifying code for pupil.
<code>sex</code>	Sex of pupil, with levels <code>female</code> and <code>male</code> .
<code>fluent</code>	Fluency at English measure taken in Year 1, where 0 = beginner, 1 = intermediate, 2 = fully fluent.
<code>ravens</code>	Test score in Year 1, marked out of 40.
<code>english</code>	English test score taken in Year 3, marked out of 100.
<code>behaviour</code>	Behaviour rating taken in Year 3, coded <code>lowerquarter</code> if pupil rated in bottom 25%, and <code>upper</code> otherwise.

Table 7: Variables in the `jspmix1` dataset, as modeled in the worked example.

and then our binary variable `behaviour`: this is modeled via a `probit` link and, since we have not specified otherwise, a constant of ones as the denominator. The default is for predictors added to the right of the tilde to have separate coefficients (i.e., one for each category) unless their name is suffixed by square brackets. In the case of the latter, a common coefficient is added to the model for the response(s) indicated in the numeric identifier within the square brackets. In this example, then, the model formula specifies separate coefficients for `sex` and `ravens`, and a common coefficient for `fluent` to be added to the response we earlier listed first (`english`). Finally, separate coefficients for the intercept are allowed to vary at level 2 (`school`) and also at level 1 (`id`), but the latter is only applicable in the case of the normal response variable (`english`; hence `1[1]`).

Next, after requesting MCMC estimation, and Gibbs sampling (denoted by 1) as the method used to update both the fixed (`fixM`) and the random effects (`residM`), and univariate MH (denoted by 2; 3 corresponds to multivariate MH – see `?write.MCMC` for further details) to update the level 1 variance (`Lev1VarM`), we specify our `runMLwiN` function call. The arguments here will be familiar to you now, although we encounter a slightly different format for the value we assign to the distribution: if our multivariate response model consisted only of normal responses, we would simply specify `"Multivariate Normal"`; since we have mixed response types, however, we must identify the distribution of each (in an order corresponding to their position in the preceding model formula), hence the character string: `D = c("Mixed", "Normal", "Binomial")`.

```

-----
MLwiN (version: 2.36) multilevel model (Mixed)
      N min      mean max N_complete min_complete mean_complete max_complete
school 47   7 23.80851 76           47             7          23.80851          76
Estimation algorithm: MCMC      Elapsed time : 3.95s
Number of obs:1119 (from total 1119) Number of iter:5000 Chains:1 Burn-in:500
Deviance statistic: NA
-----

The model formula:
c(english, probit(behaviour)) ~ 1 + sex + ravens + fluent[1] +
  (1 | school) + (1[1] | id)
Level 2: school      Level 1: id
-----

```

The fixed part estimates:

	Coef.	Std. Err.	z	Pr(> z)		[95% Cred. Interval]	ESS
sexmale_english	-6.23710	1.03073	-6.05	1.438e-09	***	-8.25154 -4.23264	4371
sexmale_behaviour	-0.41890	0.08912	-4.70	2.595e-06	***	-0.59441 -0.24209	1411
ravens_english	1.65807	0.09131	18.16	1.092e-73	***	1.47805 1.83770	3974
ravens_behaviour	0.05722	0.00774	7.39	1.456e-13	***	0.04204 0.07242	1045
fluent_1	6.36146	1.28814	4.94	7.873e-07	***	3.86164 8.85984	3720
Intercept_1	-9.19443	3.23664	-2.84	0.004501	**	-15.64483 -2.93315	3021
Intercept_2	-0.36147	0.19366	-1.87	0.06197	.	-0.74739 0.01087	1219

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

The random part estimates at the school level:

	Coef.	Std. Err.	[95% Cred. Interval]	ESS
var_Intercept_1	41.50795	12.01852	23.29346 69.47876	1734
cov_Intercept_1_Intercept_2	0.05258	0.42969	-0.77081 0.92794	628
var_Intercept_2	0.06297	0.02899	0.02167 0.13411	437

The random part estimates at the id level:

	Coef.	Std. Err.	[95% Cred. Interval]	ESS
var_bcons_2	1.00000	0.00001	1.00000 1.00000	5000
cov_bcons_2_Intercept_1	6.22663	0.70827	4.77651 7.53054	347
var_Intercept_1	291.91324	12.51061	267.92679 316.88733	740

The model indicates that the Year 1 `ravens` test score has a positive effect on both of the Year 3 response variables we have fitted, whilst the Year 1 measure of fluency in English has a positive effect on the Year 3 English test score. Girls are found to both have a higher behaviour rating, and a better Year 3 English test score, than boys. There is a positive correlation between the two response variables at the pupil-level, but the correlation is negligible at the school-level:

```
R> CompareCorrelation <- cbind(
+   MixedRespMCMC["RP"][["RP2_cov_Intercept_1_Intercept_2"]] /
+   sqrt(MixedRespMCMC["RP"][["RP2_var_Intercept_1"]] *
+   MixedRespMCMC["RP"][["RP2_var_Intercept_2"]]),
+   MixedRespMCMC["RP"][["RP1_cov_bcons_2_Intercept_1"]] /
+   sqrt(MixedRespMCMC["RP"][["RP1_var_Intercept_1"]] *
+   MixedRespMCMC["RP"][["RP1_var_bcons_2"]]))
R> colnames(CompareCorrelation) <- c("school-level", "pupil-level")
R> CompareCorrelation
```

```
      school-level pupil-level
[1,] 0.03252201 0.3644402
```

Furthermore, the model indicates that school effects explain 12.2% of the residual variation in the Year 3 English test scores, and less, 6.1%, of the residual variation in the behaviour test scores (on the latent response scale):

```
R> ExplainedBySchool <- cbind(
+   MixedRespMCMC["RP"][["RP2_var_Intercept_1"]] /
+   (MixedRespMCMC["RP"][["RP1_var_Intercept_1"]] +
+   MixedRespMCMC["RP"][["RP2_var_Intercept_1"]]) * 100,
+   MixedRespMCMC["RP"][["RP2_var_Intercept_2"]] /
+   (MixedRespMCMC["RP"][["RP1_var_bcons_2"]] +
+   MixedRespMCMC["RP"][["RP2_var_Intercept_2"]]) * 100)
R> colnames(ExplainedBySchool) <- c("english", "behaviour")
R> ExplainedBySchool
```

```
      english behaviour
[1,] 12.4491  5.923972
```

Note, as reflected in the output returned, that the DIC is currently not available for mixed response models fitted in MLwiN.

16. Running simulations/fitting multiple models

Whilst MLwiN can be run via a macro language and the command line, the R environment is particularly well-suited to requesting the estimation of a series of models, and the efficient post-processing of their results.

For example, simulation studies can be a useful tool for researchers wishing to investigate hypotheses such as how sensitive, or biased, their model estimates are to the different estimation methods which could be used to derive them. We will illustrate this with an example based on [Browne and Draper \(2000, 2006\)](#), and also Chapter 8 of [Browne \(2012\)](#), comparing Bayesian and likelihood-based methods of estimating multilevel models with very few level 2 units. Here we simulate a series of datasets from ‘true’ parameter values, and then fit each dataset to a model via the estimation methods of interest, before investigating how sensitive the model estimates are with regard to the methods used to derive them (see `demo(MCMCGuide08)` for an example of conducting such a study using parallel processing).

First we generate a dataset of 108 pupils, evenly-distributed across 6 schools; we will fit a variance components model, and so will also generate a constant (`cons`) for the intercept, and we also specify the structure of our model (the variable `resp` will be generated later):

```
R> set.seed(1)
R> pupil <- 1:108
R> school <- rep(1:6, each = 18)
R> F10 <- resp ~ 1 + (1 | school) + (1 | pupil)
```

Next we will fit two models – one via Bayesian MCMC methods and one via IGLS (accepting R2MLwiN’s default settings) – to each of 100 simulated datasets in which the response variable (`resp`) is generated anew from the pupil- and school-level variances (40 and 10, respectively) and fixed effects ($\beta_0 = 30$) for each dataset; the estimates from each model are then compiled (in `IGLS_array` and `MCMC_array`, with some additional information collected from the MCMC chains, namely the median estimate of the level 1 and levels 2 variances, and also whether the ‘true’ parameter values fall within the 95% credible (i.e., coverage) intervals):

```

R> ns <- 100
R> IGLS_array <- MCMC_array = array(, c(3, 2, ns))
R> MCMC_median <- data.frame(RP2_var_Intercept = rep(0, ns),
+   RP1_var_Intercept = rep(0, ns))
R> CounterMCMC <- rep(0, 3)
R> Actual <- c(30, 10, 40)
R> for(i in 1:ns) {
+   u_short <- rnorm(6, 0, sqrt(Actual[2]))
+   u <- rep(u_short, each = 18, len = 108)
+   e <- rnorm(108, 0, sqrt(Actual[3]))
+   resp <- Actual[1] * 1 + u + e
+   simData <- data.frame(cbind(pupil, school, resp))
+   simModelIGLS <- runMLwiN(Formula = F10, data = simData)
+   IGLS_array[, ,i] <- cbind(coef(simModelIGLS), diag(vcov(simModelIGLS)))
+   simModelMCMC <- runMLwiN(
+   Formula = F10, estoptions = list(EstM = 1), data = simData)
+   MCMC_array[, ,i] <- cbind(coef(simModelMCMC), diag(vcov(simModelMCMC)))
+   MCMC_median[i, ] <-
+   c(median(simModelMCMC["chains"][, "RP2_var_Intercept"]),
+   median(simModelMCMC["chains"][, "RP1_var_Intercept"]))
+   if (Actual[1] >
+   quantile(simModelMCMC["chains"][, "FP_Intercept"], 0.025) &
+   Actual[1] <
+   quantile(simModelMCMC["chains"][, "FP_Intercept"], 0.975)) {
+   CounterMCMC[1] <- CounterMCMC[1] + 1
+   }
+   if (Actual[2] >
+   quantile(simModelMCMC["chains"][, "RP2_var_Intercept"], 0.025) &
+   Actual[2] <
+   quantile(simModelMCMC["chains"][, "RP2_var_Intercept"], 0.975)) {
+   CounterMCMC[2] <- CounterMCMC[2] + 1
+   }
+   if (Actual[3] >
+   quantile(simModelMCMC["chains"][, "RP1_var_Intercept"], 0.025) &
+   Actual[3] <
+   quantile(simModelMCMC["chains"][, "RP1_var_Intercept"], 0.975)) {
+   CounterMCMC[3] <- CounterMCMC[3] + 1
+   }
+ }

```

Having now obtained the information we need from each estimation method, for each of the 100 simulated datasets, we now need to summarize our results; here we calculate the percentage bias of the average estimate away from the ‘true’ parameter value, together with the percentage interval coverage. For IGLS we use central Gaussian (mean $\pm 1.96 \times$ sd) intervals, but for alternative intervals see [Browne and Draper \(2006\)](#):

```

R> aa <- sapply(1:ns, function(x) na.omit(

```

```

+   stack(as.data.frame(IGLS_array[, , x]))$values)
R> counterIGLS <- rep(0,3)
R> for (i in 1:ns) {
+   if (Actual[1] > aa[1,i] - 1.96 * sqrt(aa[4,i]) &
+   Actual[1] < aa[1,i] + 1.96 * sqrt(aa[4,i])) {
+   counterIGLS[1] <- counterIGLS[1] + 1
+   }
+   if (Actual[2] > aa[2,i] - 1.96 * sqrt(aa[5,i]) &
+   Actual[2] < aa[2,i] + 1.96 * sqrt(aa[5,i])) {
+   counterIGLS[2] <- counterIGLS[2] + 1
+   }
+   if (Actual[3] > aa[3,i] - 1.96 * sqrt(aa[6,i]) &
+   Actual[3] < aa[3,i] + 1.96 * sqrt(aa[6,i])) {
+   counterIGLS[3] <- counterIGLS[3] + 1
+   }
+ }
R> Percent_interval_coverage <- (counterIGLS / ns) * 100
R> Mean_across_simus <- round(c(mean(aa[1,]), mean(aa[2,]), mean(aa[3,])), 2)
R> Percent_bias <- round(-100 * (1 - Mean_across_simus / Actual), 2)
R> IGLS_results <- cbind(Mean_across_simus, Actual, Percent_bias,
+   Percent_interval_coverage)
R> rownames(IGLS_results) <- c("beta0", "sigma2_u", "sigma2_e")
R> Percent_interval_coverage <- (CounterMCMC / ns) * 100
R> bb <- sapply(1:ns, function(x) na.omit
+   (stack(as.data.frame(MCMC_array[, , x]))$values)
R> Mean_across_simus <- round(c(mean(bb[1,]), mean(bb[2,]), mean(bb[3,])), 2)
R> Percent_bias <- round(-100 * (1 - Mean_across_simus / Actual), 2)
R> MCMC_results <- cbind(Mean_across_simus, Actual, Percent_bias,
+   Percent_interval_coverage)
R> rownames(MCMC_results) <- c("beta0", "sigma2_u", "sigma2_e")
R> cat("Simulation results using IGLS\n"); IGLS_results

```

Simulation results using IGLS

	Mean_across_simus	Actual	Percent_bias	Percent_interval_coverage
beta0	29.78	30	-0.73	87
sigma2_u	8.12	10	-18.80	77
sigma2_e	40.70	40	1.75	93

```
R> cat("Simulation results using MCMC\n"); MCMC_results
```

Simulation results using MCMC

	Mean_across_simus	Actual	Percent_bias	Percent_interval_coverage
beta0	29.57	30	-1.43	94
sigma2_u	16.20	10	62.00	91
sigma2_e	42.14	40	5.35	96

Whilst conducting fewer simulations (with fewer chain iterations too) than [Browne and Draper \(2006\)](#), the results are still largely in keeping with theirs, indicating little bias for the intercept,

slight bias (particularly via MCMC) for the level 1 variance, and considerable bias for the level 2 variance. Furthermore, the interval coverages tend to be better for MCMC, and this is especially so for the level 2 variance. Incidentally, [Browne and Draper \(2006\)](#) found that the median estimate when specifying $\Gamma(\epsilon, \epsilon)$ priors has considerably less bias, as we find here:

```
R> Mean_across_simus <- round(c(mean(MCMC_median$RP2_var_Intercept),
+   mean(MCMC_median$RP1_var_Intercept)), 2)
R> Actual <- tail(Actual, -1)
R> Percent_bias <- round(-100 * (1 - Mean_across_simus / Actual), 2)
R> Percent_interval_coverage <- tail(Percent_interval_coverage, -1)
R> MCMC_results2 <- cbind(Mean_across_simus, Actual, Percent_bias,
+   Percent_interval_coverage)
R> rownames(MCMC_results2) <- c("sigma2_u", "sigma2_e")
R> cat("Simulation results based on median MCMC estimates\n"); MCMC_results2
```

Simulation results based on median MCMC estimates

	Mean_across_simus	Actual	Percent_bias	Percent_interval_coverage
sigma2_u	10.21	10	2.10	91
sigma2_e	41.53	40	3.83	96

17. Other models and features

In addition to the models covered in the examples above, **MLwiN**, via **R2MLwiN**, has a number of other useful features; examples of many of these can be found in **R2MLwiN**'s demos (see [Section 1.3](#)), e.g., (with relevant demo in parentheses): adjusting for measurement errors in predictors ([MCMCGuide14](#)), fitting multiple membership multiple classification (MMMC) models ([MCMCGuide16](#)), fitting spatial data models ([MCMCGuide17](#)), creating imputations for multilevel datasets with missing values ([MCMCGuide18](#)), modeling autoregressive residual structures at level 1 ([MCMCGuide19](#)), and multilevel factor analysis ([MCMCGuide20](#)).

18. Conclusions

This paper has introduced **R2MLwiN**, a new package which calls the multilevel modeling software **MLwiN** from within the R environment. As such, it offers a route other than the GUI, or direct authoring of macros, to run **MLwiN**, enabling users to benefit from working within the R environment (e.g., to parsimoniously fit multiple models, and compile and analyze the results returned using the many statistical and graphical functions available in R). In addition, **R2MLwiN** offers functionality not commonly-supported by existing R packages, such as modeling complex level 1 variance, multiple membership models, multivariate response models, writing (and running) **BUGS** models with automatically-generated model code and **IGLS** starting values, and **MLwiN**'s interactive equation and graphics windows available via its GUI, which can be opened from R (if running via **Windows** or in **Wine** ([Wine Development Team 2015](#)) on other platforms). As a means to call **MLwiN** externally **R2MLwiN** joins the recently-developed programme **Stat-JR** ([Charlton et al. 2013](#)) (which interoperates with a large range of statistical software programmes) and the **Stata** package ([StataCorp. 2015](#))

runmlwin (Leckie and Charlton 2013). This work therefore has the potential to be of benefit to a range of research communities, offering an accessible means of addressing a common research problem: namely modeling datasets with multilevel structures.

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